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Full Length Research Paper

Molecular Epidemiology of *Escherichia coli* in Raw Cow's Milk: Prevalence and Genetic Determinants of Virulence

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Raw milk plays an important role in the survival and transport of pathogenic bacteria including *Escherichia coli* strains. This study was performed to determine the existence of *E. coli* in raw milk intended for human consumption and its associated virulence determinants. In this context, a total of 232 milk samples were obtained from different dairy shops located at Mansoura city and its surrounding villages. Milk samples were subjected for bacteriological and serological examination of *E. coli*. Furthermore, *E. coli* strains were tested for its haemolytic activity on blood agar plates. The recovered *E. coli* strains were also screened by Polymerase chain reaction (PCR) for the presence of enterotoxins including heat –labile (LT), heat- stable (ST) toxins and haemolysin (*hly*) genes. The recovery rate of *E. coli* was 14.65% (34/232). Among the recovered *E. coli* strains, 12 different *E. coli* serotypes were identified namely, O26:H11 (n=6), O111:H2 (n=5), O128:H2 (n=5), O91:H21 (n=4), O124 (n=3), O127:H6 (n=3), O103:H21 (n=2), O153 (n=1), O113:H4 (n=2), O6 (n=1), O121:H7 (n=1) and O146 (n=1). Regarding PCR results, 31(91.19%) *E. coli* strains harbored *STa* and seven strains carried *hly* gene (20.59%) while non *E. coli* isoates harbord *LT* gene. Conclusively, raw milk can be considered as serious source of pathogenic *E. coli*, therefore, proper management practices and effective control measures are recommended to improve milk hygiene and sanitation.

Key words: Raw milk, *Escherichia coli*, enterotoxin genes, haemolysin gene.

INTRODUCTION

Raw milk harbor variable microorganisms, considered as an important source of food borne pathogens because it is regarded as perfect media for microbial growth (Laba and Udosek, 2013). Consumption of raw milk may be associated with the occurrence of food-poisoning outbreaks (Christidis et al., 2016). The presence of

different food borne pathogens in milk may be contributed to the fecal contamination during milking process (Rehman et al., 2014). *E. coli* is a normal inhabitant of the gastrointestinal tract of both man and animals. Most of *E. coli* strains are harmless, but some are known to be pathogenic bacteria, causing severe intestinal and extra

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Table 1. Sequences and cycling conditions of oligonucleotide primers.

Primer	Sequence	Amplified product	Reference	Primary denaturation	Secondary denaturation	Annealing	Extension	No. of cycles	Final extension
hly	AACAAGGATAAGCACTGTTCTGGCT ACCATATAAGCGGTCATTCCCGTCA	1177 bp	Piva et al. (2003)	94°C 5 min	94°C 30 s	60°C 1 min.	72°C 1 min	35	72°C 12 min
STa	GAAACAACATGACGGGAGGT GCACAGGCAGGATTACAACA	229 bp	Lee et al.	94°C 5 min	94°C 30 s.	57°C 30 s.	72°C 30 s	35	72°C 7 min
Lt	GGTTTCTGCGTTAGGTGGAA GGGACTTCGACCTGAAATGT	605 bp	(2008)	94°C 5 min	94°C 30 sec	57°C 45 s	72°C 45 s	35	72°C 10 min

intestinal diseases in man (Croxen et al., 2013). Isolation of E. colifrom milk represent a serious public health hazard because some strains of E .coli may be belongs to enteropathogenic or toxigenic or both groups which causes sever gastrointestinal disturbance (Thomas et al., 2017). Enterotoxigenic E. coli (ETEC) is one of the most common bacteria responsible for diarrhea in different parts of the world (Bagheri et al., 2014). Like other gastrointestinal infectious diseases, they are caused by lack of sanitation and most often contamination transfers from contaminated food or water (Walker et al., 2007; Marchou, 2013). There are two enterotoxins, Heat-stable toxin (ST) and Heat-labile toxin (LT). These two toxins are considered as the main virulence factors which influence the pathogenesis of ETEC strains (Kolenda et al., 2015; Sjöling et al., 2015). Alpha-hemolysin (HlyA) of *E. coli* is one of cytolytic pore-forming toxins (PFTs) produced by Gram-negative bacteria. E. coli HlyA lyses erythrocytes shows strong cytotoxic and cytolytic action against diversity of nucleated cells (Söderström et al., 2017). HlyA does not only kill and lyse cells but also affects target cells at sublytic concentrations. Haemolysin (hlyA) is produced mainly by extraintestinal pathogenic E. coli (ExPEC) strains and occasionally by ETEC, STEC and EPEC (Burgos and Beutin, 2010). Therefore, the main purpose of this study was to examine E. coli isolated from raw milk for the presence of enterotoxins including heat labile (LT) and heat stable (ST) toxins and haemolysin.

MATERIALS AND METHODS

Sampling

A total of 232 raw milk samples were collected randomly from different dairy shops, groceries and supermarkets in Mansoura city and its surrounding villages at Dakhalia Governorates, Egypt during the period from January to April, 2017. All samples were collected in sterile tubes and transported in an ice box to the laboratory as quick as possible for bacteriological examination with minimal of delay.

Isolation and identification of E. coli

All samples were immediately centrifuged and the sediment were

streaked onto the surface of MacConkey's agar plates and incubated aerobically at 37°C for 24 h (Quinn et al., 2002). Lactose fermenting (Pink colored) colonies was sub-cultured on Eosin Methylene Blue (Oxoid) agar medium.

Colonies showing characteristic metallic green sheen on EMB agar were identified as *E. coli*. Presumptive *E. coli* colonies were subjected for gram staining and standard biochemical tests (Quinn et al., 2004). Additional identification of *E. coli* isolates was performed using commercial biochemical test kits (bioMerieux API, France).

Serological identification of *E. coli*

E. coli strains were transferred to Food Analysis Center, Faculty of Veterinary Medicine, Benha University for serological identification using rapid diagnostic *E. coli* antisera sets (Kok et al., 1996).

Haemolytic activity

E. coli isolates were cultured on blood agar containing 5% sheep blood, for detection of its haemolytic activity. Haemolysis was recorded after an overnight incubation at 37°C. A clear halo was defined as haemolysin positive (Brauner et al., 1990).

PCR assay for detection of enterotoxin genes (STa-LT) and haemolysin gene (*hly*)

Bacterial genomic DNA was extracted from *E. coli* isolates according to Ramadan et al. (2016). *E. coli* isolates were screened by Polymerase chain reaction (PCR) for the presence of enterotoxins (*Lt, STa*) and haemolysin (*hly*) genes. Oligonucleotide primers sequences and its amplicons sizes were described in Table1. Amplification reaction of PCR targeting haemolysin and enterotoxins was performed as previously described by Piva et al. (2003) and Lee et al. (2008), respectively (Table 1). Amplified DNA products for each gene were analyzed by 1.5% agarose gel electrophoresis in 1x TBE buffer stained with ethidium bromide visualized under UV transillumator.

RESULTS

In the present study, 34 (14.65%) *E. coli* strains have been recovered out of 232 examined milk samples. Among *E. coli* strains, twelve different *E. coli* serotypes were identified including, O26, O111, O128, O91, O124,

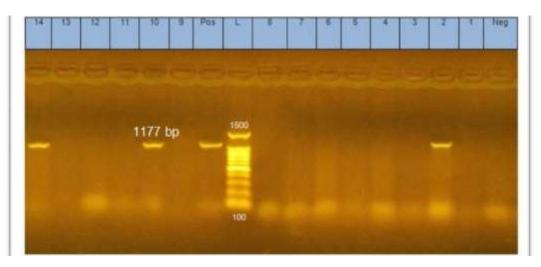


Figure 1. Agarose gel electrophoresis demonstrating amplification of *hly* gene at 1177 bp. Pos: Positive control, L: 100 bp DNA ladder, Neg: Negative control.

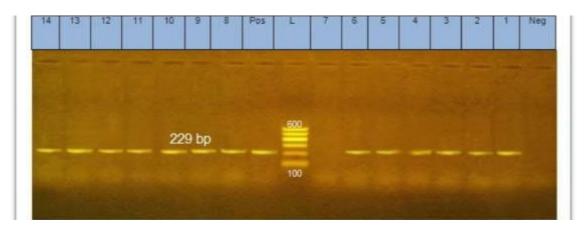


Figure 2. Agarose gel electrophoresis demonstrating amplification of STa gene at 229bp. Pos: Positive control, L: 100 bp DNA ladder, Neg: Negative control.

O127, O103, O113, O153, O6, O121 and O146 with a prevalence rate of 17.6, 14.7, 14.7, 11.7, 8.8, 8.8, 5.8, 5.8, 2.9, 2.9, 2.9 and 2.9% respectively.

E. coli isolates were tested for hemolytic activity on 5% sheep blood agar, 52.94% (18/ 34) of E. coli strains revealed different degrees of hemolysis. Based on the PCR results, 91.19% of E. coli isolates are potentially pathogenic, which carry one or more investigated virulence genes. From a total of 34 E. coli strains, 7(20.59%) strains carried hly gene (Figure 1), 31(91.17%) strains carried STa (Figure 2) while none E. coli isolates carried LT gene (Table 2).

DISCUSSION

Raw milk is a perfect medium that supports the growth and multiplication of *E. coli*. Consumption of such milk

appeared as main threat to health concerns, especially for those people who still drink raw milk without heat treatment (Claeys et al., 2013). In the present study, *E.coli* was recovered with 14.65% prevalence rate. Similarly, *E. coli* has been isolated by several researchers from raw milk of cattle and buffaloes (Caine et al., 2014; Islam et al., 2008; Hossain et al., 2011). Compering to present results, a higher percentage of *E. coli* in milk was reported by Bandyopadhyay et al. (2012), Farzan et al. (2012), Mohd et al. (2013), Ali and Abdelgadir (2011) and Gwida and EL-Gohary (2013), who could isolate *E. coli* from raw milk in a percentage of 26.43,30.28, 33.96, 63 and 41.2% respectively. However, lower results were recorded by Kivaria et al. (2006) who detected *E. coli* in 6.3% of the examined raw milk samples.

In the present study, 12 different *E.coli* serotypes were identified; nearly the same serotypes were recovered

Table 2. Prevalence of serotypes,	Enterotoxin (ST	and LT) and	haemolysin genes (hly)
of E. coli isolated from raw milk.			

Caretura (Na)	Number (9/)	hly gone (7)	Enterotoxin genes		
Serotype (No)	Number (%)	hly gene (7)	LT	STa	
O26:H11	6(6.71)	3	-	6	
O111:H2	5(6.7.)	-	-	5	
O91:H21	4 (667.)	-	-	4	
O103:H21	2(875)	-	-	2	
O113:H4	2(875)	-	-	-	
O153	1(972)	1	-	1	
O6	1(972)	1	-	1	
O121:H7	1(2.9)	1	-	1	
O146	1(2.9)	1	-	-	
O124	3(8.8)	-	-	3	
O127:H6	3(8.8)	-	-	3	
O128:H2	5(14.7)	-	-	5	
Total	34	7	-	31	

from raw milk samples by Helmy et al. (2011) and Osman et al. (2012). Hemolysin is one of the important virulent factors in E. coli. In this study, 52.94% E. coli isolates revealed hemolysis on 5% sheep blood agar. Similarly, Farzan et al. (2012) reported that, one E. coli strain out of three isolates showed \(\beta \)-hemolytic activity on blood agar also, Lamey et al.(2013) found that 12.7% of isolated E. coli strains were hemolytic, Sayed (2014) found that one isolate (5.6%) out of 18 E. coli isolates had hemolytic activity. Concerning hly gene, 20.59% of E. coli harbored hly gene. A lower percentage was recorded by Ombarak et al.(2016), who identify hly gene in 2 (2.25%) isolates from karish cheese and one isolate (0.90%) from raw milk while, a higher percentages (42.85%) were reported by Osman et al. (2012). The presence of E. coli in milk especially enteropathogenic and/or toxigenic strains has a public health hazards which lead to sever gastrointestinal disturbance. Among E. coli isolates, 7(20.59%) strains carried hly gene, 31(91.17%) strains carried STa while, LT gene was not identified in all E. coli strains. Comparing to these results. Eid (2014) revealed that, only one strain were tested positive for STa gene out of eight E. coli isolates.

In Brazil, Paneto et al. (2007) studied the frequency of toxigenic *E. coli* in raw milk and cheese whereby, 1(2%) of *E. coli* isolates were ETEC. In Romania, Tabaran et al. (2017) analyzed 145 *E. coli* strains isolated from raw milk and traditional dairy cheeses, for the presence of *STa* and *STb*. In *LT*, none of the samples carries the *estl* gene, but 14 (9.7%) of the *E. coli* isolates were positive for both *eltA* and *estll*. Caine et al. (2014) examined 100 *E. coli* strain for the presence of enterotoxins which could identify enterotoxins in 4% of the total examined isolates.

Bonyadian et al. (2014) tested 120 isolates of *E. coli*, isolated from milk and unpasteurized cheeses which

identified LT and STb in 2(1.66%) and 12(10.00%), respectively but could not identify *STa* gene. In this study, it was interesting that, all *E. coli* strains carry *hly* gene along with enterotoxin gene. These results suggest that food of animal origin represents a significant source of pathogenic *E. coli* strains.

Conclusion

The high contamination of milk with toxigenic *E. coli* represents a serious public health hazards which necessity high and strict preventive measures, to minimize the bacterial contamination within the food chain such as regular washing and sterilization of dairy equipment, utensils, milker's hands, animal udders as well as heat treatment of milk before distribution to consumers.

Significance statement

This study provided a data about the prevalence of *E. coli* in cow's raw milk, especially enterotoxigenic (ETEC) *E. coli*. These data is required for the establishment of food control systems which required the prevention and control of foodborne illnesses.

CONFLICT OF INTERESTS

The authors have not declared any conflict of interests.

REFERENCES

Ali AA, Abdelghadir SW (2011).Incidence of *Escherichia coli* in raw cow's milk in Khartoum state.Br J. Dairy sci. 2(1):23-26.

- Bagheri S, Mousavi Gargari SL, Rasooli I, Nazarian S, Alerasol M (2014).
 A CssA, CssB and LTB chimeric protein induces protection against Enterotoxigenic Escherichia coli. Braz. J. Infect. Dis. 18(3):308-314.
- Bandyopadhyay S, Lodh C, Rahaman H, Bhattacharya D, Bera AK, Ahmed FA, Mahanti A, Samanta I, Mondal DK, Bandyopadhyay S, Sarkar S, Dutta TK, Maity S, Paul V, Ghosh MK, Sarkar M, Baruah KK (2012).Characterization of shiga toxin producing (STEC) and enteropathogenic *Escherichia coli* (EPEC) in raw yak Poephagusgrunniens) milk and milk products. Res. Vet. Sci. 93(2):604-610.
- Bonyadian M, Bonyadian M, Moshtaghi H, Taheri MA (2014). Molecular characterization and Antibiotic resistance of enterotoxigenic and entero-aggregative *Escherichia coli* isolated from raw milk and unpasteurized cheeses. Vet. Res. Forum. 5(1):29-34.
- Brauner A, Katouli M, Tullus K, Jacobson SH (1990). Production of cytotoxic necrotizing factor, verocytotoxin and haemolysin by pyelonephritogenic *Escherichia coli*. Eur. J. Clin. Microbial Infect. Dis. 6:762-767.
- BurgosY, Beutin L (2010). Common origin of plasmid encoded alphahemolysin genes in *Escherichia coli*. BMC Microbiol. 10:193.
- Caine LA, Nwodo UU, Okoh AI, Ndip RN, Green E (2014). Occurrence of Virulence Genes Associated with Diarrheagenic *Escherichia coli* Isolated from Raw Cow's Milk from Two Commercial Dairy Farms in the Eastern Cape Province, South Africa. Int. J. Environ. Res. Public Health. 11(11):11950-11963.
- Christidis T, Pintar KD, Butler AJ, Nesbitt A, Thomas MK, Marshall B, Pollari F (2016). Campylobacter spp. Prevalence in Raw Milk: A Systematic Review and Meta-Analysis. J. Food Prot. 79 (10):1775-1783.
 - Claeys WL, Cardoen S, Daube G, Block JD, Dewettinck K, Katelijne Dierick K, Zutter LD, Huyghebaert A, Imberechts H, Thiange P, Vandenplas Y, LieveHerman L (2013). Herman Raw or heated cow milk consumption: Review of risks and benefits. Food Contl. 31:251-262
- Croxen MA, Law RJ, Scholz R, Keeney KM, Wlodarska M, Finlay BB (2013). Recent advances in understanding enteric pathogenic *Escherichia coli*. Clin. Microbiol. Rev. 26:822-880.
- Eid AM (2014). Molecular identification of some contagious microorganisms causing food poisoning from bulk tank milk in Gharbia Governorate. Benha Vet. Med. J. 27(2):29-47.
- Farzan R, Rahimi E, Momtaz H (2012). Virulence properties of Shiga Toxin-Producing *Escherichia coli* isolated from Iranian raw milk and dairy products. Slov. Vet. Res. 49(4):159-66.
- Gwida, MM, El-Gohary FA (2013). Zoonotic bacterial pathogens isolated from raw milk with special reference to *Escherichia coli* and *Staphylococcus aureus*. Governorate, Egypt. Open Access Sci. Rep. 2(4):705-708.
- Helmy SM, Ammar MA, Aisha RA, El-Shabrawy MA, Hakim AS, Bakry MA, Abuelnaga, AS, Eraqi MM (2011). Molecular and virulence characterization of *Escherichia coli* strains isolated from persistent bovine mastitis. J. Am. Sci. 7:614-624.
- Hossain Z, Sultana P, Deb S, Ahmed MM (2011). Multidrug resistance in large-plasmid-associated presumptive enterohaemorrhagic *Escherichia coli* isolated from contaminated lake water. Bangladesh J. Microbiol. 28:33-40.
- Islam MA, Mondol AS, de Boer E, Beumer RR, Zwietering MH, Talukder KA, Heuvelink, AE (2008). Prevalence and genetic characterization of shiga toxin-producing *Escherichia coli* isolates from slaughtered animals in Bangladesh. Appl. Environ. Microbiol. 74:5414-5421.
- Kivaria FM, Noordhuizen JPTM, Japaga AM (2006). Evaluation of the hygienic quality and associated public health hazards of raw milk marketed by small holder dairy producers in the Dar es Salaam region, Tanzania. Trop. Anim. Health Prod. 38(3):185-194.
- Kok T, worswich D, Gowans E (1996). Some serological techniques for microbial and viral infections. In practical Medical Microbiology (collee, J.; Fraser ,A.; Marmion, B. and simmons A., eds.) 14thed., Edinburg, Churchill Livingstone, UK.
- Kolenda R, Burdukiewicz M, Schierack P (2015). A systematic review and meta-analysis of the epidemiology of pathogenic *Escherichia coli* of calves and the role of calves as reservoirs for human pathogenic *E. coli*. Front Cell Infect. Microbiol.5:23.

- Laba SA, Udosek CE (2013). Bacteriological quality of raw cow milk in Ilorin, north central Nigeria. Nat. Sci. 11(10):73-79.
- Lamey AE, Ammar AM, Zaki ER, Khairy N, Moshref BF, Refai MK (2013). Virulence factors of *Escherichia coli* isolated from recurrent cases of clinical and subclinical mastitis in buffaloes. Int. J. 4(1):86-94.
- Lee SI, Kang SG, Kang ML Yoo HS (2008). Development of multiplex polymerase chain reaction assays for detecting enterotoxigenic *Escherichia coli* and their application to field isolates from piglets with diarrhea. J. Vet. Diagn. Invest. 20:492-496.
- Marchou B (2013). Traveller's diarrhea: epidemiology, clinical practice guideline for the prevention and treatment. Presse Med. 42(1):76-81.
- Mohd R, Kotwal SK, Malik MA, Singh M (2013). Prevalence, genetic profile of virulence determinants and multidrug resistance of *Escherichia coli* isolates from foods of animal origin. Vet World. 6:139.
- Ombarak RS, Hinenoya A, Awasthi Atsushi, SP, Iguchi A, Shima, A, Elbagory AM (2016). Yamasaki S. Prevalence and pathogenic potential of *Escherichia coli* isolates from raw milk and raw milk cheese in Egypt. Int. J. Food Microbiol. 221:69-76.
- Osman KM, Mustafta KM, Aly AK AbdElhamed GS (2012). Serotypes, virulence genes, and intimin types of Shiga toxin-producing *Escherichia coli* and enteropathogenic *Escherichia coli* isolated from mastitic milk relevant to human health in Egypt. Vector Borne Zoonotic Dis. 12(4):297-305.
- Paneto BR, Schocken-Iturrino RP, Macedo C, Santo E, Marin JM (2007).

 Occurrence of toxigenic *Escherichia coli* in raw milk cheese in Brazil.

 Arq. Bras. Med Vet. Zootec. 59:508-512.
- Piva IC, Pereira AL, Ferraz LR, Silva RSN, Vieira AC, Blanco JE, Blanco M, Blanco J. Giugliano, LG (2003). Virulence Markers of Enteroaggregative *Escherichia coli* Isolated from Children and Adults with Diarrhea in Brasília, Brazil. J. Clin. Microbiol. 41(5):1827-1832.
- Quinn PJ, Markey BK, Carter ME, Donnelly WJ Leonard FC (2002). Veterinary Microbiology and Microbial Diseases. 2ed. Blackwell Science Ltd. NdBodmin, Cornwall, UK.
- Quinn J, Carter M E, Markey B Carter GR (2004). Clinical veterinary microbiology. 6th ed., Mosby, Edinburgh, London, New York, Philadelphia, St. Louis, Sydney, Toronto.
- Ramadan H, Awad A, Ateya A (2016). Detection of phenotypes, virulence genes and phylotypes of avian pathogenic and human diarrheagenic *Escherichia coli* in Egypt. J. Infect. Dev. Ctries. 10(6):584-591.
- Rehman MU, Rashid M, Sheikh JA, Bhat, MA (2014). Molecular epidemiology and antibiotic resistance pattern of enteropathogenic *Escherichia coli* isolated from bovines and their handlers in Jammu. India. J. Adv. Vet. Anim. Res. 1(4):177-181.
- Sayed SM (2014). A contribution on Coliforms causing mastitis in cows with reference to serotypes and virulence factors of *E. coli* isolates. Ass. Univ. Bull. Environ. Res. 17(1):85-95.
- Sjöling A, Mentzer, von A, Svennerholm AM (2015). Implications of enterotoxigenic *Escherichia coli* genomics for vaccine development. Expert. Rev. Vaccines 14 (4):551-560.
- Söderström CM, Fagerberg SK, Brogaard MB, Leipziger J, Skals M, Praetorius HA (2017). Loop Diuretics Diminish Hemolysis Induced by α-Hemolysin from *Escherichia coli*. J. Membr .Biol. 250(3):301-313
- Tabaran A, Mihaiu M, Tăbăran F, Colobatiu L, Reget O, Borzan M.M, Dan SD (2017). First study on characterization of virulence and antibiotic resistance genes in verotoxigenic and enterotoxigenic *E. coli* isolated from raw milk and unpasteurized traditional cheeses in Romania. Folia. Microbiol. (Praha). 62(2):145-150.
- Thomas RR, Brooks HJ, O'Brien R (2017). Prevalence of Shiga toxin-producing and enteropathogenic *Escherichia coli* marker genes in diarrhoeic stools in a New Zealand catchment area. J. Clin. Pathol. 70(1):81-84.
- Walker RI, Steele D, Aguado T (2007). Analysis of strategies to successfully vaccinate infants in developing countries against enterotoxigenic *E. coli* (ETEC) disease. Vaccine 25(14):2545-2566.