

Full Length Research Paper

# Mitigative Effects of Calcium on Cadmium-Induced Oxidative Stress and Anti-Oxidative Enzyme Activity in Tomato Plants

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In the present study, the cadmium's effect on anti-oxidative enzymes in tomato (*Lycopersicon esculentum*) plants was evaluated. Plants treated with increasingly CdCl<sub>2</sub> concentrations (0, 20, 40, 80, 100 and 200 µM respectively) were grown in a basic nutrient solution for 7 days. APX (Ascorbate Peroxidase) and GPX (Guaïgol Peroxidase) activities showed to increase below 100 µM concentration after treatment. In excessive concentration however, more than determined level, a significant decrease in the enzymes activities was determined. The increase in enzymatic activity can be associated with induction of oxidative stress by cadmium treatment. Excessive calcium displayed also low APX and GPX activities, which were virtually the same activity levels of control plants. However, it should be note that concentration more than 80 µm of Cd resulted in decreasing of APX and GPX activity. The enzymatic activity of GSH followed in the presence of Cd showed a significant increase by 80 µM of Cd then the activity showed decreasing. Calcium treatment resulted in impairing of GSH stimulation in the presence of Cd conditions. Eventually, Cd treatment resulted in decline in GST activity that was significantly increased in the presence of calcium, suggesting stimulation of antioxidant enzymatic activities. Therefore, accumulation of Cd can be associated with oxidative stress, which may be inhibited and reduced its adverse effects by calcium treatment in the cell culture.

**Keywords:** Cadmium, Calcium, APX, GPX, GSH, GST, *Lycopersicon esculentum*, enzymatic activity.

## INTRODUCTION

Many parts of the world, especially near urban and Industrial areas, are highly polluted by heavy metals resulted from human activity. Among all these heavy metals: Cd, Cu, Hg, Ni, Pb and Zn are the most dangerous. Now the use of mineral fertilizers, pesticides, sludge and sewage are among the main sources of soil contamination by cadmium (Wagner, 1993; Ravera, 2001).

Cadmium is relatively rare in the ecosphere (McBride, 1995). It is one of the most toxic pollutants and more mobile in the soil-plant system. Therefore, its assimilation and accumulation in the tissues of plants can be animal or human source of contamination by consumption Grant and Bailey (1995). Indeed, the Cadmium is an oxydore- ductor toxic heavy metal not known to interact with living organisms. It can increase the formation of intracellular

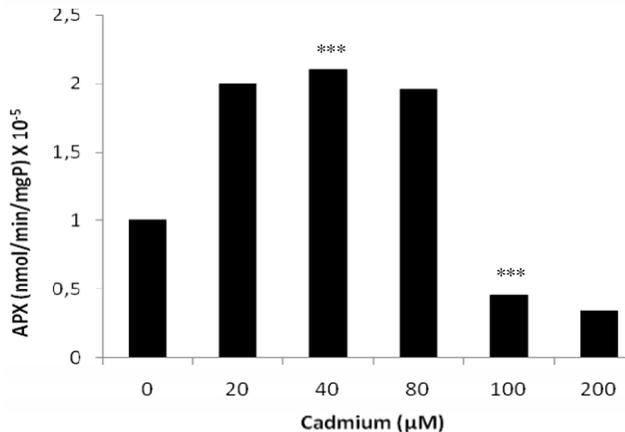
Reactive Oxygen Species (ROS) and generates an oxidative stress (Lagadic et al., 1997).

As a defence mechanism, the plant cells involve in enzymatic system consisting special antioxidants such as α-tocopherol, β-carotene, Glutathione-S-Transferase (GSH) and ascorbate. In addition to other antioxidant enzymes, Superoxide Dismutase (SOD), Ascorbate Peroxidase (APX), Catalase (CAT) and Glutathione reductase (GR), were involved in the eradication of reactive species with oxygen against this toxicity (Rosen, 2002).

Tripeptide glutathione consists of glutamate cysteine and glycine; it plays a crucial role in defending against free radicals (AOS) in plants that are under the oxidative stress conditions (De Vos et al., 1992; Ranieri et al., 1993). It is also, the precursor of phytochelatin that hold heavy metals in plants (Rosen, 2002).

In this study, we investigated in the first hand the behaviour of plants treated with a xenobiotic (cadmium) compound, and the antioxidative protection system components in which GST (Glutathione-S-Transferase)

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**Figure 1.** APX Activity of Tomato roots treated with different concentrations of Cd ( $p < 0.001$ ).

reduced glutathione (GSH), Ascorbate Peroxidase (APX) and Guaïgol-peroxidase (GPX) activities that have been associated with the possible role of calcium against cadmium in tomato (*Lycopersicon esculentum*) (Kauss, 1987; Hagemeyer and Breckle, 1996; Kinraide, 1998).

## MATERIAL AND METHODS

### Conditions of culture

The tomato seeds (*L. esculentum*, Rio Grandy Variety) were disinfected with a solution of peroxide hydrogen at 10% (V/V), rinsed thoroughly with distilled water, and then were placed into Petri plates and the sprout were grown on filter paper soaked in distilled water.

Germination was done in the dark at  $25 \pm 1^\circ\text{C}$ . Seedlings older than 7 days were transplanted into nutrient solutions (Zoghlimi-Boulila et al., 2006). pH was between 5.5 and 6.5. The composition of the culture medium was  $\text{KNO}_3$ : 100.1 g/l;  $\text{KH}_2\text{PO}_4$ : 136.09 g/l; Citrate iron: 38.91 g/l;  $\text{ZnSO}_4$ : 0.28 g/l;  $\text{H}_3\text{BO}_3$ : 1.85 g/l;  $\text{MgSO}_4$ : 246.4 g/l;  $\text{CaCl}_2$ : 147 g/l;  $\text{CuSO}_4$ : 0.25 g/l;  $\text{MnSO}_4$ : 0.84 g/l.

After 12 days in culture, the seedlings were transferred in the same medium consisting  $\text{CdCl}_2$  in different doses (0, 20, 40, 80, 100 and 200  $\mu\text{M}$ ) or control without  $\text{CdCl}_2$  for 7 days. In order to investigate cadmium-calcium interaction, calcium  $\text{CaCl}_2$  (1mM) was added to the basic solutions containing different concentrations of  $\text{CdCl}_2$ .

The different nutritional mediums were continuously aerated with a bubbling compressed air pump under the hood in Laboratory.

### Analytical techniques

#### Enzymatic extraction

Tomato roots were extracted according to Loggini (1999). The extract was used for the estimation of the activity of ascorbate peroxidase (APX), Guaicol-peroxidase (GPX), and Glutathione-S- Transferase (GST).

At 12 days of treatment, 1 gram of roots sample was cooled, and crushed cold in a mortar with 5ml of phosphate buffer (pH 7.5). The homogenate was properly filtered before making any cold centrifuge  $4^\circ\text{C}$  at 12000 g for 20 min (Sigma 3-16K). The obtained supernatant was used as the extract for the determination of the dif-

ferent enzymatic activities.

#### Determination of APX

Ascorbate Peroxidase activity was measured according to the protocol of Azada and Nakano (1987). The final reaction volume of 3ml contains 100 $\mu\text{l}$  of enzyme extract, 50  $\mu\text{l}$  of  $\text{H}_2\text{O}_2$  in 0.3% and 2.85 ml of NaK-ascorbate buffer (50 mM of NaK, 0.5 mM of ascorbate, pH7.2). The absorbance was measured by spectrophotometer at 290 nm for 1min, for a coefficient of linear molar extinction  $\epsilon = 2800 \text{ M}^{-1} \cdot \text{cm}^{-1}$ . The APX activity was expressed as nmol/min/mg of protein.

#### Determination of GPX

The guaïacol-peroxidase (GPX) activity is determined by colorimetric method (spectrophotometer Jenway 6300) at 470 nm (Chaoui et al., 1997) the coefficient of linear molar extinction is  $\epsilon = 2470 \text{ M}^{-1} \cdot \text{cm}^{-1}$ . The final volume of 3 ml contains 100  $\mu\text{l}$  of enzyme extract, 50  $\mu\text{l}$  of  $\text{H}_2\text{O}_2$  in 0.03% and 2.85 ml of phosphate-Guaïacol buffer (50 mM of NaK, 8 mM of guaïacol, pH 7.2). The reaction was started by the addition of peroxide hydrogen. GPX activity is expressed as nmol/min/mg of protein.

#### Determination of GSH

The enzyme extract was homogenized in a solution of 0.02 M EDTA, then, it undergoes a disinfecting, moistening by sulfo-Sali-cyclic acid 0.25%. After centrifugation at 2000 g for 10min, supernatant was used for the spectrophotometric assay at 412 nm with the reagent DTNB at 0.01 million. The GSH activity was expressed in  $\mu\text{M}/\text{mg}$  of protein (Anderson, 1985).

#### Determination of the GST and total protein

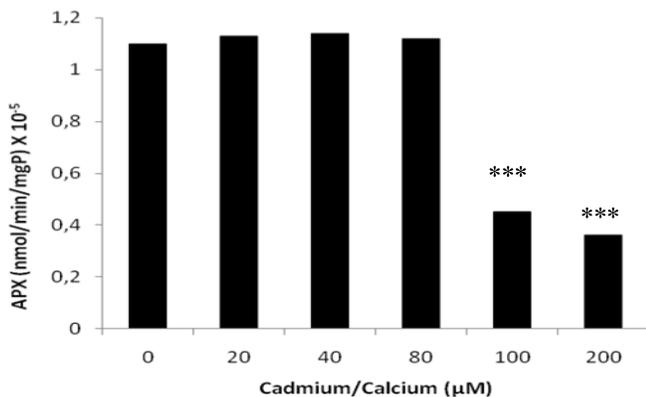
Glutathione-S-Transferase level was estimated using Habig (1974) method. Samples were homogenized in a 100 mM of phosphate buffer in pH 6.5, and then it was centrifuged at 9000 g for 30 min. The method was based on the reaction of the GSTs in a mixture of CDNB (20 mM) and GSH (100 mM), the change in optical density is due to the emergence of complex CDNB-GSH that measured every 15 seconds for 2 min at 340 nm. Concentration of the GST was expressed in nmol/min/mg of protein. The total protein level was measured according to Bradford (1976) method.

## RESULTS AND DISCUSSION

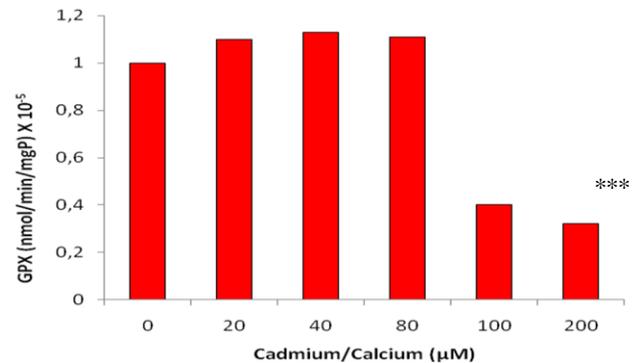
In despite of their positive role in several metabolic processes, heavy metals cause severe cellular damage (Kyunghee and Junghoon, 2001). The molecular mechanisms by which cells defend against stress generated by heavy metals are an important focus of research.

The results show a strong stimulation of the APX and GPX enzymatic activity in treated plants roots with low concentrations of Cd (Figure 1 and 3), this result shows accordance with the results of Chaoui et al. (1997).

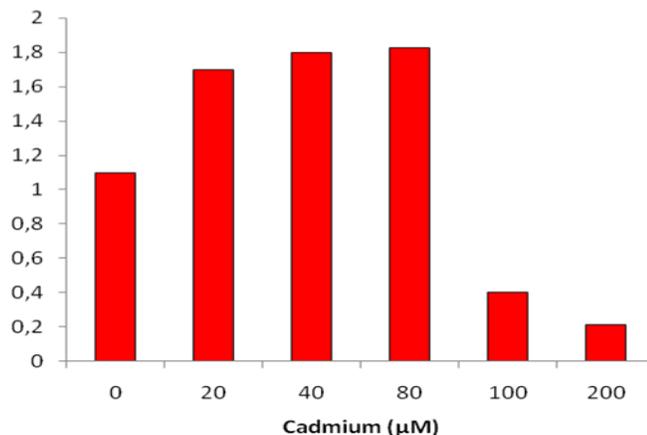
Meanwhile, the increased activity of GSH were observed in our study (Figure 5), suggesting the involvement of this enzyme in the detoxification of (RSO) and free radicals (Asada and Takahashi, 1987). Moreover, compared to enzymatic activity recorded in control plants



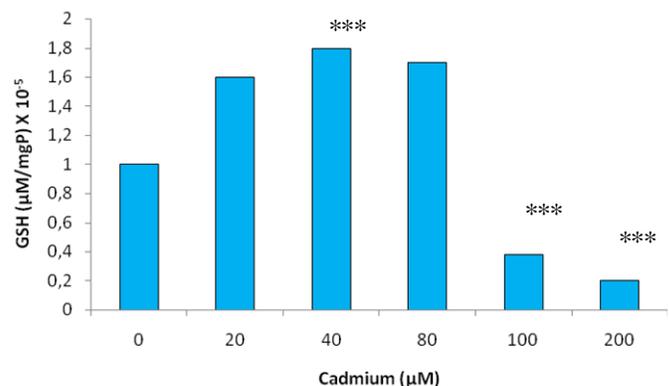
**Figure 2.** APX Activity of tomato roots treated with different concentrations of Cd and combined with Ca (Cd/Ca) ( $p>0.05$ ), with 100 and 200  $\mu\text{M}$  ( $p<0.001$ ).



**Figure 4.** GPX Activity of Tomato roots treated with different concentrations of Cd and combined with Ca (Cd/Ca) ( $p>0.05$ ).



**Figure 3.** GPX activity of tomato roots treated with Cd ( $p<0.001$ ).



**Figure 5.** GSH Activity of Tomato roots treated with Cd ( $p<0.001$ ).

and those processed by the low concentrations of Cd, the obtained results in treatment with 100 and 200  $\mu\text{M}$  of Cd clearly put in evidence the toxic effect of this metal. This is fully consistent with the reported observations on *Brassica* by Zhu et al. (1999), *Pteris vitata* by Cao et al. (2004) and *Tilapia caerulea* by Freeman et al. (2004), where the standard of GSH is directly linked with increased tolerance to the accumulation of Cd at concentrations below 100  $\mu\text{M}$ .

Likewise, the obtained results on *Scenedesmus biju-gatus* by Prasad et al. (1999) and *Helianthus annuus* by Gallego et al. (2002) where the level of GSH decreases in response to stress produced by Cd and Cu at concentrations above 100  $\mu\text{M}$ .

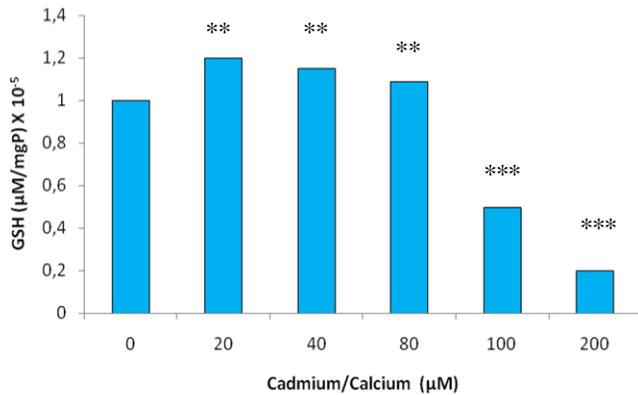
Variations of registered GST show that cadmium tends to stimulate this activity slightly at low concentrations (20 to 80  $\mu\text{M}$ ). Beyond these concentrations the toxic effect of this xenobiotic is clearly established (Figure 7). This result clearly highlights the role of this enzyme in the tolerance

of tomato plants to cadmium.

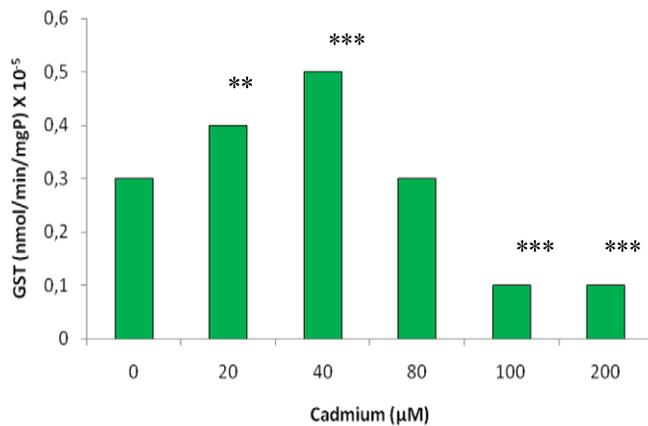
The addition of 1mM of calcium raises significantly the inhibitory effect observed in the presence of cadmium for all measured enzymes (Figure 2, 4, 6 and 8). This effect is due to directly entering of the calcium ion in competition with cadmium for the same sites thereby blocking the influx of the metal within the cells. These results are compared with those obtained by Bernal and Ruvalcaba (1995) and Fellenberg (2000).

In conclusion, the results suggest that treatment with low concentrations of cadmium (under 100  $\mu\text{M}$ ) generates hydrogen Peroxide and low ROS accumulation (Un-Haing and Nam-Ho, 2004). This low accumulation may boost a range of defence systems in cell damage, these systems may lead to stimulation of antioxidant enzymatic activities (APX, GPX, GSH, GST), playing a decisive role in the detoxification of cells which are involved in (Chaffei et al., 2003).

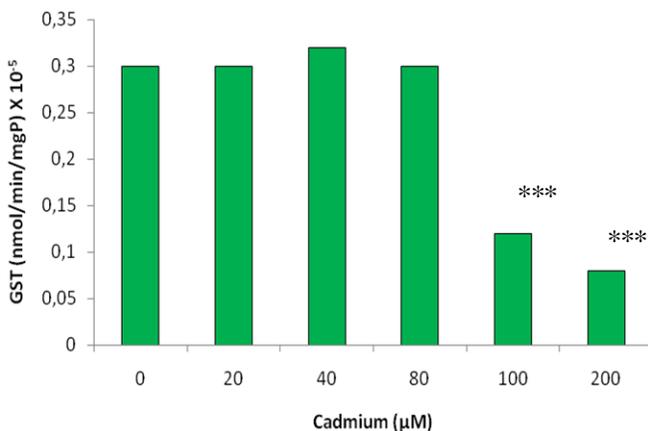
The combined treatment Cd/Ca showed to measured activity level of the enzymes as in the cells of control plants. The addition of calcium lift the inhibition of enzymatic activity induced by cadmium treatment was as suggested in previous studies (Parameswaran and Majeti, 2003; Romero-Puertas et al., 2007).



**Figure 6.** GSH Activity of Tomato roots treated with different concentrations of Cd and combined with Ca (Cd/Ca) ( $p < 0.01$ ).



**Figure 7.** GST Activity of Tomato roots treated with Cd ( $p < 0.001$ ).



**Figure 8.** GST Activity of Tomato roots treated with different concentrations of Cd and combined with Ca (Cd/Ca) ( $p < 0.001$ ).

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