

Full Length Research Paper

# Genetic characterization of three breeds of high royal jelly producing honeybee (*Apis mellifera ligustica*) in China

Ling Yin<sup>1</sup>, Ting Ji<sup>1\*</sup>, Guohong Chen<sup>1</sup> and Wenjun Peng<sup>2</sup>

<sup>1</sup>College of Animal Science and Technology, Yangzhou University, Yangzhou 225009, China.

<sup>2</sup>Institute of Honeybee, Chinese Academy of Agricultural Sciences, Beijing 100094, China.

Accepted 24 November, 2017

There are three most famous breeds of high royal jelly producing honeybee (*Apis mellifera ligustica*) in China. After numerous rounds of selection for royal jelly production, the royal jelly production of the three breeds is much higher than the unselected. It is important to maintain genetic diversity of breeds in selective breeding and stock management. The genetic diversity and genetic differentiation of three breeds were surveyed using 18 microsatellite markers. Using Polymorphism Information Content (PIC), mean heterozygosity (H), number of effective alleles, genetic distances, gene flow (Nm) and F-statistics, we evaluated the genetic diversity and genetic differentiation. The result showed that the number of alleles per locus ranged from 2 (AP156 and A028) to 25 (AP053). All of the three breeds showed high levels of heterozygosity. Significant genetic differentiation was found among the three breeds and the average genetic differentiation coefficient of the three breeds was 0.037.

**Key words:** High royal jelly producing honeybee, *Apis mellifera ligustica*, genetic diversity, genetic differentiation, microsatellite marker.

## INTRODUCTION

Since the consumer demand on royal jelly in Asia is much high, many honeybee breeding programs were constructed to obtain high royal jelly producing breeds in China. After several years of selection, the royal jelly production of honeybee *Apis mellifera ligustica* in China has improved a lot, and there are three most famous breeds (Xiaoshan bee, Pinghu bee, Zhenongda NO.1) cultivated in Zhejiang province, which are used widely around the country. Maintenance of genetic diversity of breeds is an important factor in selective breeding and stock management. Reductions in diversity promote susceptibility to disease outbreaks and other negative conditions associated with inbreeding (Baer and Schmid-Hempel, 2001; Tary and Seeley, 2006; Seeley and Tary, 2007). To maintain the achievement of the

breeding programs, the genetic characterization of three breeds should be assessed.

With the characteristics of high polymorphism, locus specificity, abundance and random distribution over the genome, and their co-dominant inheritance, microsatellites are currently the most commonly used to assess population structure and diversity (Chapman, 2008; Bourgeois and Rinderer, 2009; Delaney et al., 2009; Zarkti et al., 2010). According to FAO recommendations, determining classic genetic distances using neutral, highly polymorphic microsatellite markers is the method of choice for investigating genetic relationships and breed differentiation. This methodology also provides information for establishing preservation priorities for livestock breeds (Barker, 1999).

The aim of this research was to evaluate genetic diversity and genetic differentiation of the 3 high royal jelly producing breeds with 18 microsatellite markers. The results may be useful to understand genetic Characterization of the three breeds and contribute to a

\*Corresponding author. E-mail: [jiting12@sohu.com](mailto:jiting12@sohu.com).  
Tel: 86-13305275665. Fax: 86-514-7342351.

**Table 1.** The location of 18 microsatellite loci in chromosome and PCR conditions.

	GenBank accession NO.	Chromosome	Mg <sup>2</sup> concentration (mmol/L)	Annealing temperature
AG005a	AJ509722	Chr LG1	2	55
AC306	AJ509721	Chr LG2	2	55.6
AP274	AJ509486	Chr LG3	2	55
AP043	AJ509329	Chr LG3	2	56.5
AP313	AJ509504	Chr LG4	2	57
AP053	AJ509338	Chr LG5	2.2	56.5
AP143	AJ509400	Chr LG5	2.2	56.5
A113	AJ509290	Chr LG6	2	58.2
A014	AJ509239	Chr LG8	2	55.6
AC011	AJ509637	Chr LG9	1.8	57
AP189	AJ509433	Chr LG10	1.6	57
AP156	AJ509410	Chr LG10	2	55.6
BI299	BI514528	Chr LG11	2	55.6
AP085	AJ509359	Chr LG12	2	55.6
AT101	AJ509549	Chr LG12	2	55.6
AT003	AJ509505	Chr LG13	2	55.6
A028	AJ509244	Chr LG14	2	55
AP068	AJ509351	Chr LG15	2	55.6

more efficient selection and breeding.

## MATERIALS AND METHODS

### Sampling

A total of 144 individual bees, 48 worker bees of each breed were analysed in this study. Xiaoshan bees (XS) were collected from Bee Breeding Farm in Xiaoshan, Zhejiang and Pinghu bees (PH) were collected from Bee Breeding Farm in Pinghu, Zhejiang and Zhenongda NO.1 (ZN) were collected from Zhejiang University.

### DNA isolation

DNA was extracted from the thorax of individual bees (one bee per colony) according to the method reported by Ji et al. (2005).

### Genotyping

The DNA polymorphism was assessed at 18 microsatellite loci (Table 1). These markers are randomly distributed across 14 of the 16 chromosomes of *Apis mellifera* genome. The primers are selected according to NCBI (<http://www.ncbi.nlm.nih.gov>). PCR products were obtained in a 20 l volume using thermal cycler. Each PCR tube contained 50 ng of genomic DNA, 2.0 l of 10xbuffer 1.2 to 2.0 l of 25 mmol/l MgCl<sub>2</sub> 0.5 l of 10 mmol/l dNTP, 1 l of both 10 pmol/l forward primer and reverse primer, 5 U/l Taq DNA Polymerase 0.2 l. The amplification involved initial denaturation at 95°C (5 min), 35 cycles of denaturation at 95°C (5 s), annealing temperature varying between 50 and 60°C (50 s),

and extension at 72°C (50 s), followed by final extension at 72°C (10 min). DNA fragments were scored on 8% polyacrylamide gel using a ABI 377 automated DNA analyzer (P-Eapplied Biosystem, America). Electrophoregram processing was performed with GENESCAN3.1 software (P-Eapplied Biosystem, America), and allele-size scoring was analyzed by the Binthere software (P-EApplied Biosystem, America).

### Statistical analysis

**Genetic diversity:** Total number of alleles, allele frequencies, average number of alleles per locus, observed (Ho) and expected heterozygosity (He) for each population across the loci, were estimated with Microsatellite-Toolkit for Excel (Park, 2001).

**Genetic differentiation:** Population differentiation was estimated by Wright's (1978) fixation indices  $F_{IT}$ ,  $F_{ST}$  and  $F_{IS}$  in the form of  $F$ ,  $f$ , and  $f$ , respectively, for each locus across populations according to the variance based method of Weir and Cockerham (1984) using FSTAT software (Version 2.9.3, Goudet, 2002). The significance of the F-statistics was determined by permutation tests with the sequential Bonferroni procedure applied over loci (Hochberg, 1988). The extent of inbreeding was further studied with GENEPOP software (Raymond and Rousset, 1995) by estimating the  $F_{IS}$  values and their significance level within each of the populations.

Pair-wise  $F_{ST}$  values were computed for all combinations of the 3 populations using GENEPOP software. Gene flow between populations, defined as the number of reproductively successful migrants per generation ( $Nm$ ), was estimated based on the  $n$  island model of population structure (Slatkin and Barton, 1989). The estimate was based on the relationship  $F_{ST} = 1 / (4Nm + 1)$ , where  $N$  is the effective population size,  $m$  is the migration rate, and  $F_{ST}$  is calculated as mean over loci. The Reynolds' genetic distance (Reynolds et al., 1983) between breeds was calculated, based on

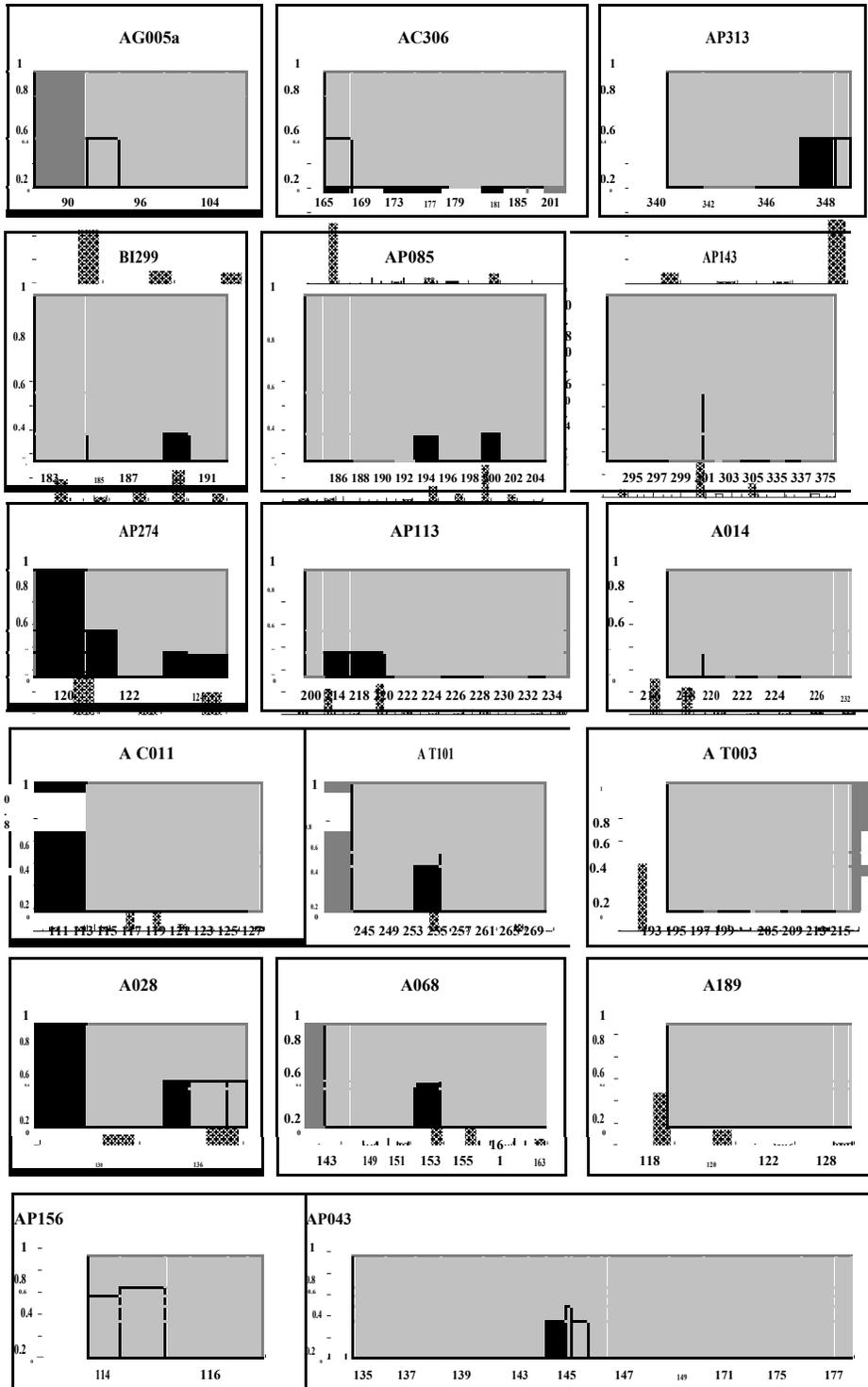


Figure 1. Graphs of microsatellite allele frequencies of 18 loci used to assess genetic diversity of 3 high royal jelly producing honeybees breeds. ■, XS; □,PH; and▨, ZN.

$F_{ST}$  values.

## RESULTS

### Genetic variability within populations

In total, 18 microsatellite loci were used for polymorphism

analyses among the three breeds (Table 1). A total of 135 alleles were detected in the three breeds with varying allele frequencies by the 18 microsatellite markers (Figure 1). Among the three breeds, the number of alleles was consistent (Xiaoshan bee, 103; Pinghu bee, 107 and Zhenongda NO.1, 110). The alleles for each breed overlapped but were not identical. Unique alleles were evident within each breed (XS = 8, pH = 9, and ZN = 12).

**Table 2.** Total number of alleles, expected heterozygosity (He), Observed heterozygosity (Ho) and PIC.

Locus	No. of alleles	(Ho)	(He)	PIC
AG005a	3	0.529	0.583	0.467
AC306	8	0.420	0.382	0.397
AP274	3	0.577	0.458	0.499
AP043	10	0.660	0.688	0.630
AP313	4	0.393	0.451	0.330
AP053	25	0.943	0.847	0.937
AP143	9	0.341	0.361	0.325
A113	11	0.688	0.653	0.641
A014	7	0.553	0.444	0.472
AC011	9	0.782	0.813	0.748
AP189	4	0.487	0.451	0.419
AP156	2	0.214	0.174	0.191
BI299	5	0.693	0.701	0.645
AP085	10	0.756	0.764	0.719
AT101	8	0.551	0.583	0.483
AT003	8	0.196	0.188	0.192
A028	2	0.193	0.174	0.174
AP068	7	0.613	0.667	0.571
Mean	7.500 (5.2497)	0.533 (0.2100)	0.521 (0.2133)	0.491 (0.2083)

Standard deviations for mean number of alleles, He and PIC, were given in parentheses.

**Table 3.** Mean number of alleles per locus and the average heterozygosity (He and Ho) for three breeds of *Apis mellifera ligustica*.

Breed	Number of alleles (mean ±SD)	Average He (Mean ± SD.)	Average Ho (mean ±SD)
XS	5.722±4.586	0.519±0.057	0.536±0.017
PH	5.944±4.518	0.522±0.048	0.507±0.017
ZN	6.111±4.391	0.518±0.053	0.521±0.017
Total	7.500±5.250	0.533±0.050	0.521±0.010

Allelic richness differed between breeds but was highly varied among loci (Tables 2 and 3).

Expected heterozygosity (He) and mean polymorphic information content (PIC) for each locus across three breeds were listed in Table 2. The average number of the alleles observed in 18 microsatellite loci was 7.50. Across the three breeds, locus A028 had the lowest He, 0.193 and the lowest PIC, 0.174, however, the locus AP053 had the highest He and PIC value, 0.943 and 0.937, respectively.

### Genetic differentiation among populations

The average number of alleles per locus expected and observed heterozygosity and  $F_{IS}$  for each breed across 18

loci were shown in Table 3. Zhenongda NO. 1 had the highest value of average number of alleles per locus, 6.111 and Xiaoshan bee had the lowest one with 5.722, the value of Pinhu bee was 5.944. The estimates of expected heterozygosity of the three breeds were much higher and similar to each other (XS, 0.519; PH, 0.522; ZN, 0.518). The lowest value of observed heterozygosity (0.507) was obtained for Pinhu breed, while the highest one (0.536) was found in Xiaoshan breed.

The fixation indices ( $F_{IT}$ ,  $F_{ST}$ ,  $F_{IS}$ ) for each locus across all populations are shown in Table 4. The fixation coefficients of subpopulations within the total population, measured as  $F_{ST}$  value, for the 18 loci varied from -0.005 (AC306) to 0.284 (AT101), with a mean of 0.037 ( $P<0.001$ ). 13 of 18 loci contributed significantly to this differentiation. The global deficit of heterozygotes across populations ( $F_{IT}$ ) amounted to 0.034 ( $P<0.01$ ). The negative  $F_{IS}$  values of some loci indicated an excess of heterozygous genotypes with respect to the expected value. Mean  $F_{IS}$  was found to be -0.004 within populations. Four loci showed significant of excess heterozygotes, while no marker showed significant of deficit heterozygotes.

Population genetic structure measures showed no evidence ( $P > 0.05$ ) for inbreeding but did show significant levels of diversity among breeds (Figure 2). The three breeds were differentiated significantly from each other, Xiaoshan bee differed from Pinghu bee and

**Table 4.** The results from F-statistics analysis.

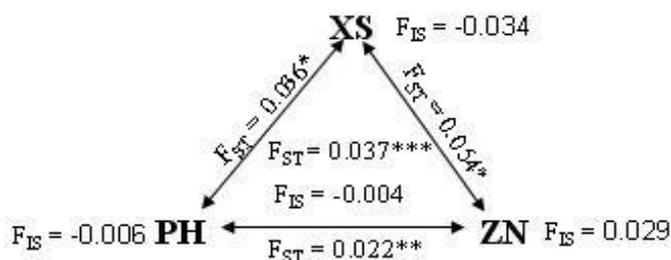
Locus	$F_{IT} = F$	$F_{ST} = \theta$	$F_{IS} = f$
AG005a	-0.101	0.005	-0.106
AC306	0.089	-0.005	0.094
AP274	0.213***	0.024***	0.194**
AP043	-0.023	0.053***	-0.08
AP313	-0.139	0.025*	-0.168
AP053	0.106***	0.013***	0.094***
AP143	-0.036	0.067***	-0.111
A113	0.054	0.008**	0.047
A014	0.213***	0.059***	0.164*
AC011	-0.039	-0.001**	-0.039
AP189	0.091	0.057***	0.036
AP156	0.196**	0.02	0.179*
BI299	-0.01	0.005	-0.015
AP085	-0.005	0.018***	-0.023
AT101	0.039	0.284***	-0.341
AT003	0.048	0.01*	0.038
A028	0.105	0.017	0.089
AP068	-0.087	0.004**	-0.092
Mean	0.034**(0.024)	0.037***(0.017)	-0.004(0.029)

Mean estimates from jack-knife over loci, standard deviations are given in parentheses; \* $p < 0.05$   
 \*\* $p < 0.01$  \*\*\* $p < 0.001$ .

**Table 5.** Reynolds' genetic distances,  $D_R$  (upper triangle) and the gene flow,  $N_m$  (lower triangle) between three breeds.

Breed	XS	PH	ZN
XS		0.037	0.055
PH	6.714		0.022
ZN	4.405	11.011	

Estimated of gene flow ( $N_m$ ) and Reynolds' genetic distances ( $D_R$ ) between each population pair are presented in Table 5. The lowest values of Reynolds' distance was 0.022 (Pinghu bee-Zhejiangnongda NO.1 pair), and the highest one was 0.055 (Xiaoshan bee-Zhejiangnongda NO.1 pair). The  $N_m$  value ranged from 4.405 (between Xiaoshan bee- Zhejiangnongda NO.1 pair) to 11.011 (between Pinghu bee-Zhejiangnongda NO.1 pair).



**Figure 2.** Graphical representation of three breeds. Genetic diversity estimates are listed along each axis. Values inside of figure represent overall diversity estimates. \* $P < 0.05$ . \*\* $P < 0.01$ .

Zhenongda NO.1 significantly ( $F_{ST} = 0.036$ ,  $P = 0.013$ ;  $F_{ST} = 0.054$ ,  $P = 0.034$ , respectively), and Pinghu bee differed from Zhenongda NO.1 more significantly ( $F_{ST} = 0.002$ ,  $P = 0.009$ ).

## DISCUSSION

### Genetic variability within breeds

18 microsatellite markers used in the present study are randomly distributed across 14 chromosomes in the *Apis mellifera* genome, so the data had certain comparability and representativeness. The polymorphism information content (PIC) value is a good measure of the polymorphisms of gene fragment, while  $PIC > 0.5$ , the locus is a highly polymorphic locus; while  $0.25 < PIC < 0.5$ , the locus is a medium polymorphic locus; while  $PIC < 0.25$ , the locus is a low polymorphic locus (Vanhalala et al., 1998). Meanwhile, PIC value is related to the availability and utilization efficiency of a marker, the higher PIC value of the marker, the higher heterozygote frequency in one population, as well as the more genetic information it provides. In this study, 11 loci among 18

microsatellite loci exhibited high polymorphic, while 4 loci showed medium polymorphic, mean PIC value across all loci exceeded 0.5, which could provide enough information for the assessment of genetic diversity.

Number of alleles is also good for measuring the genetic variation, especially in conservation genetics study. Sometimes its effect on populations is put more emphasis, but effective number of alleles is easy to be affected by sample size (Maudet et al., 2002). The average number of the alleles was 7.50 across 18 microsatellite loci in the present study, which indicated that the sample size was enough. On the other hand, this result also indicated that the polymorphism information content provided by these 18 microsatellite loci in the three breeds was rich; and the distribution of the allelic frequency was rather even.

Gene heterozygosity, also called gene diversity, is a suitable parameter for investigating genetic variation. Ott (2001) gave a definition that a polymorphic locus must have at least 0.10 heterozygosity. Mean expected heterozygosity can approximately reflect the variation of genetic structure. All 18 microsatellite loci in this study had high polymorphism with a mean expected heterozygosity, 0.533, showing a high degree of genetic diversity. Overall levels of gene diversity, and allelic richness were high although little lower compared with levels found in a microsatellite survey of commercial Italian bee populations in the United States and Italy (Bourgeois et al., 2008), Russian honey bee stock selected for improved resistance to *Varroa destructor* (Bourgeois and Rinderer, 2009) and commercial populations surveyed in Western Australia (Chapman et al., 2008).

### Genetic differentiation among breeds

In our study, on average, the genetic differentiation ( $F_{ST}$ ) among breeds was 3.7% (Table 4), a relative high value and extremely significant ( $P < 0.001$ ), which indicated that there is a great differentiation among the three breeds. It is clear that about 3.7% of the total genetic variation corresponds to differences of breeds and the remaining 96.3% is the result of differences among individuals. Most of the loci contribute to this differentiation significantly. And the three breeds were differentiated significantly from each other (Figure 2). The values of  $N_m$  and Reynolds' genetic distances between pairs of breeds also supported the differentiation of the three breeds.

The coefficient  $F_{IS}$ , which indicates the degree of departure from random mating, positive  $F_{IS}$  values mean a significant deficit of heterozygotes, while the negative  $F_{IS}$  values indicate an excess of heterozygous genotypes with respect to the expected value. In this study negative average of  $F_{IS}$  was -0.004, but not significant. In addition, four loci (AP274, AP053, A014 and AP156) showed significant excess of heterozygotes.

Many honeybee breeding programs were constructed to obtain high royal jelly producing breeds in China, and three most famous breeds Xiaoshan bee, Pinghu bee, Zhenongda NO.1 were cultivated in Zhejiang province. The estimation of characteristics of the three breeds indicated that the three breeds had been successful in maintaining heterozygosity and high levels of diversity while keeping inbreeding levels at a minimum. Genetic diversity measures should be monitored to ensure that heterozygosity and allelic richness are maintained at their current levels.

### ACKNOWLEDGEMENTS

This work was performed in College of Animal Science and Technology, Yangzhou University, China. The authors would like to thank Xiaoshan Bee Breeding Farm, Pinghu Bee Breeding Farm and Zhejiang University for providing honey bee samples. This project was supported by grants NYCYTX-43 from Special Fund for Modern Bee Industry and Technology System provided by China Agriculture Ministry.

### REFERENCES

- Baer B, Schmid-Hempel P (2001). Unexpected consequences of polyandry for parasitism and fitness in the bumblebee, *Bombus terrestris*. *Evolution*, 55: 1639-1643.
- Barker JSF (1999). Conservation of livestock breed diversity. *Anim. Genet. Res. Inf.*, 25: 33-43.
- Bourgeois AL, Danka SR, Rinderer T (2008). Comparison of microsatellite DNA diversity among commercial queen breeder stocks of Italian honey bees in the United States and Italy. *J. Apic. Res.*, 47: 93-98.
- Bourgeois AL, Rinderer TE (2009). Genetic Characterization of Russian Honey Bee Stock Selected for Improved Resistance to *Varroa destructor*. *J. Econ. Entomol.*, 3: 1233-1238.
- Chapman NC, Lim J, Oldroyd BP (2008). Population Genetics of Commercial and Feral Honey Bees in Western Australia. *J. Econ. Entomol.*, 2: 272-277.
- Delaney DA, Meixner MD, Schiff NM, Sheppard WS (2009). Genetic Characterization of Commercial Honey Bee (Hymenoptera: Apidae) Populations in the United States by Using Mitochondrial and Microsatellite Markers. *Ann. Entomol. Soc. Am.*, 4: 666-673.
- Goudet J (2002). FSTAT version 2.9.3.2. Department of Ecology and Evolution, University of Lausanne, LAUSANNE, Switzerland.
- Hochberg Y (1988). A sharper Bonferroni procedure for multiple test of significance. *Biometrika*, 75: 800-802.
- Ji T, Chen J, Pan R (2005). Compare of bee's genome DNA concentration by different abstracting method. *Apiculture of China*, 12: 10-11.
- Maudet C, Miller C, Bassano B (2002). Microsatellite DNA and recent statistical methods in wild conservation management: Application in Alpine ibex *Capra ibex (ibex)*. *Mol. Ecol.*, 11: 421-436.
- Ott J (2001). *Analysis of Human Genetic Linkage* (revised edition). Baltimore: Johns Hopkins University Press.
- Park SDE (2001). Trypanotolerance in West African Cattle and the Population Genetic Effects of Selection (Ph.D. thesis). University of Dublin.
- Raymond M, Rousset F (1995). GENEPOP (version 1.2): Population genetics software for exact test and ecumenicism. *J. Heredity* 86:248-249.
- Reynolds J, Weir BS and Cockerham CC (1983). Estimation of the coancestry coefficient: basis for a short-term genetics distance. *Genetics*, 105: 767-769.

- Seeley TD, Tarpay DR (2007). Queen promiscuity lowers disease within honeybee colonies. *Proc. Biol. Sci. R. Soc.*, 274: 67-72.
- Slatkin M, Barton NH (1989). A comparison of three indirect methods of estimating average levels of gene flow. *Evolution*, 43: 1349-1368.
- Tarpay DR, Seeley TD (2006). Lower disease infections in honeybee (*Apis mellifera*) colonies headed by polyandrous vs monandrous queens. *Naturwissenschaften*, 93: 195-199.
- Vanhala T, Tuiskula-Haavisto M, Elo K (1998). Evaluation of genetic distances between eight chicken lines using microsatellite markers. *Poult. Sci.*, 77: 783-790.
- Weir BS, Cockerham CC (1984). Estimation F-statistics for the analysis of population structure. *Evolution*, 38: 1358-1370.
- Wright S (1978). *Evolution and the genetics of populations-Variability within and among natural populations*. Vol. 4. University of Chicago press, Chicago, IL, USA.
- Zarkti H, Ouabbou H , Hilali A, Udupa SM (2010). Detection of genetic diversity in Moroccan durum wheat accessions using agromorphological traits and microsatellite markers. *Afr. J. Agric. Res.*, 14: 1837-1844.