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Full Length Research Paper

Factors influencing the sero-prevalence of *Trypanosoma brucei gambiense* sleeping sickness in Juba District, Central Equatoria State, Southern Sudan

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A surveillance of the Gambian type of Human African Trypanosomosis (HAT) conducted in Juba area using the Card Agglutination Test for Trypanosomosis (CATT) showed that 257 (11.1%) out of 2322 individuals were sero-positive. The sero-positive rate in the pooled adults was higher but not significanty different from that of the pooled children. The adult females sero- prevalence rate was significantly higher than those of adult males, male or female children. There were no significant differences in the sero-prevalence rates between adult men and male or female children. The internally-displaced group and the military personnel showed statistically higher sero-positive rates than the resident groups regardless of the presence or absence of the only extant tsetse species, *Glossina fuscipes fuscipes*. The proportions of sero-positives differed significantly between locations in the study area. The respondents examined along the riverine vegetation had a statistically higher encounter with the disease than those in the derived savanna and the open savanna woodland. There was no significant correlation between the density of *G. fuscipes fuscipes* and the incidence of *Trypanosoma brucei gambiense*.

Key words: CATT, sero-prevalence, sleeping sickness, HAT, Gambian type.

INTRODUCTION

At the end of the19th century, sleeping sickness decimated the populations of vast areas in the Equatoria region of Southern Sudan (Kuzoe, 1991). The devastating epidemic in Southern Sudan was attributed to the Gambian form of the disease (Bloss, 1960; Ford, 1971) although, Sudan lies in the edge of the geographical distribution of the rhodesiense and gambiense, the two

Abbreviations: HAT, Human African trypanosomosis; **CATT,** card agglutination test for trypanosomosis.

types of sleeping sickness. Additionally, at that time there were no effective diagnostic techniques, which might have lead to the suspicion that the causative agent was anything other than *Trypanosoma brucei gambiense*. The main characteristic feature of the history of Human African Trypanosomosis (HAT) in the Sudan is a cycling between periodic devastating epidemics interspersed with long periods of low-level endemicity.

The epidemics of sleeping sickness flared up mainly as a sequel to civil unrest in the country or its neighbours, the Congo and Uganda, most likely due to population dislocation and the collapse of the health system it caused (Hunt and Bloss, 1945; Bloss, 1960; Hutchinson, 1975; Snow, 1983; 1984; Kuzoe, 1993; Moore et al., 1999; Moore and Richer, 2001).

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Figure 1. Study area showing the various location surveyed for tsetse and Gampian sleeping sickness in Bahr El Jebel State, Sudan.

Presently, the national efforts are expanding to address the problems posed by sleeping sickness in Southern Sudan. Yet, there are serious limiting factors that restrict and suspend these efforts. These include security and instability altogether with movement of refugees and the internally-displaced people which have led to new epidemiological patterns that are inadequately elucidated.

However, the recent introduction of Card Agglutination Trypanosomosis Test/ *T. brucei gambiense* (CATT), a serodiagnostic screening test for antibodies detection, during active surveillance has been a major breakthrough in the control of sleeping sickness (CATT/wb: Magnus et al.,1978). The present HAT surveillance was conducted using CATT in the accessible villages around metropolitan Juba, the Central Equatoria State capital, together with the accessible camps of the military and internallydispalced people.

The main objective was to identify the factors influceing the sero- prevalence of sleeping sickness in the country in order to understand the disease epidemiology, and consequently to formulate a suitable integrated strategy for control.

MATERIALS AND METHODS

Locations surveyed

The study was conducted in Juba district (Latitude 4° 40'-5° N and

Longitude 30° 30'-31° 45' E). The areas covered represented the major types of vegetation. In the derived savanna woodland west of Juba town the locations examined were: Nyamini, Koda, Serimon, El -Jebalain and Rokon, in the open savanna woodland North west of Juba, the locations were: Kapu, and Luri, in the riverine vegetation South of Juba on the western bank of River Bahr El Jebel, Lologo, Khor Rommalla, Tokiman, Rejaf West, Nyori and on the eastern bank of the river: Gumbo, Rejaf East, Khor Kit and Logo East, were screened (Figure 1).

Screening procedures for human African trypanosomosis

At each location visited, at least 95% of the consenting inhabitants were screened for *T. brucei gambiense* sleeping sickness using the card agglutination test for trypanosomosis (CATT/wb: Magnus et al., 1978). Blood was collected from the basilic veins of consenting individuals using heparinized capillary tubes. Confirmed CATT positives were recorded according to location, vegetation, residence, gender, age group and tsetse occurrence.

Tsetse survey

The biconical trap is effective for sampling *Glossina palpalis* group tsetse (Challier et al., 1977; Odulaja and Mohamed-Ahmed, 1997) and the Epsilon trap is similarly effective against *Glossina morsitans* group tsetse (Hargrove and Langley, 1990). Since the two groups of tsetse flies are thought to occur in the study area both of the latter traps were used in order to define the tsetse occurrence, their diversity and apparent densities. In each vegetation type traps were placed 200 m apart for 3 days during each of the wet and dry seasons. Captured flies (pooled males and females) were collected

Classification of individuals	% sero-prevalence	Chi ² value	Probability, P
Pooled adults	11.9 (194/1630)	3 6df – 1	p = 0.058
Pooled children	9.10(63/692)	5.001 - 1	p = 0.000
Declad males	0 50 (400/4450)		
Pooled males	9.53 (139/1459)	9.05df = 1	p<0.0026
Pooled females	13.67 (118/863)		
Dealed a duit we also	0 45 (404/4400)		
Pooled adult males	9.45 (104/1100)	18.61df = 1	p<0.00001
Pooled adult females	16.98 (90/530)		F
Dealed a duit we also	0 45 (404/4400)		
Pooled adult males	9.45 (104/1100)	3.80df = 1	p>0.9
Pooled male children	9.75 (35/359)		F
Pooled adult females	16.98 (90/530)	12.02 df = 1	p<0.0005
Pooled female children	8.41 (28/333)		Protocoo
	0.75(05(050)		
Pooled male children	9.75(35/359)	0.23df = 1	p>0.63
Pooled female children	8.41(28/333)	0.2001 - 1	F: 5100

Table 1. The percentage of inhabitants seropositive for human African trypanosomosis according to age groups and gender in Juba area, Central Equatoria State, Sudan.

every 24 h, identified, sexed and counted. It is a fact that the transmission of *T. b. gambiense* occurs among humans is through the agency of tsetse flies, the *palpalis* group. For this reason we tried to find out the relationship between the density of tsetse (x) in each location and the seroprevalence rate (y) in the same location using the simple linear regression equation (y = a + bx) where a is the intercept or constant and (b) is the regfression coefficient or slope.

RESULTS

Human African trypanosomosis (HAT) seroprevalence

The surveillance of HAT, covered 2322 persons. Regardless of age and sex, 257 individuals (11.07%) of the total scanned were seropositive.

Sero-prevalence of HAT based on pooled gender and age-groups

The results are presented in Table 1. The sero-prevalence rate in the pooled adults, although, are relatively higher than in the pooled children and the difference was marginally insignificant (Chi² = 3.6; p = 0.058). However, regression analysis showed a significant positive correlation between the square of age of respondents regardless of sex or location (x) and their HAT seroprvalence rate (y) (Y = 1.07 + 0.039x; r = 0.9656; df =169; t = 2.082; p < 0.04). On the other hand, the adult females sero-prevalence rate was significantly higher than that of the adult males, male or female children (Chi² = 9.05; df = 2, p = 0.0026). In contrast, there was no significant difference in the sero-prevalence rate between men and male children.

Sero-prevalence of HAT based on residence

The pooled internally-displaced group showed a statistically higher HAT seropositive rate (p < 0.000), when compared with the pooled resident groups where *Glossina fuscipes fuscipes* had been caught in biconical traps or the pooled resident groups where tsetse had not been seen in traps. Conversely, paired comparisons showed insignificant difference in HAT encounter between the two resident groups (Chi² = 0.007:p > 0.93) (Table 2).

HAT sero- prevalence based on gender and age group in relation to residence

Table 3 compares the sero-prevalence rates in the internally-displaced respondents according to gender and age group. Paired comparisons showed significant the variations between pairs excluding male children/female children. Furthermore, comparisons between residents of tsetse infested areas showed essentially similar sero-prevalence rates irrespective of sex or age (Table 4). However, for the inhabitants of tsetse free areas there were significant discrepancies in sero-prevalence rates between sexes and age groups barring adult female/female children and male children/ female children (Table 5). Subsequent regression

Classification of individuals	% sero-prevalence	Chi ² value	Probability, P
Total displaced	18.09 (131/724)	52.75	P<0.000
Total resident with tsetse	7.93 (23/290)	df = 2	
Total resident with no tsetse	7.87 (103/1308)		
Total displaced	18.09 (131/724)	15.82	P<0.0007
Total resident with tsetse	7.93 (23/290)	df = 1	
Total displaced	18.09 (131/724)	46.76	P<0.000
Total resident with no tsetse	7.87 (103/1308)	df = 1	
Total resident with tsetse	7.939 (23/290)	0.007	P>0.93
Total resident with no tsetse	7.87 (103/1308)	df = 1	

Table 2. The percentage of male and female inhabitants seropositive for Human African Trypanosomosis according to residence in Juba area, Central Equatoria State, Sudan.

Table 3. The percentage of internally- displaced people seropositive for human African trypanosomosis according to gender and age-groups in Juba area, Central Equatoria State, Sudan.

Classification of individuals % sero-prevalence		Chi ² value	Probability, P	
Pooled adults	26.93(108/401)	29.46 df - 1	n <0.000	
Pooled children	9.16(23/251)	20.40 ul = 1	ρ<0.000	
Pooled adult males	21.97 (49/223)			
Pooled adult females	49.58 (59/178)	5.724 df = 1	p<0.0167	
Pooled adult males	21 97 (19/223)			
Pooled male children	9.09(11/121)	8.167 df = 1	p<0.0042	
Pooled adult females	49.58 (59/178)	22.89 df = 1	p<0.000	
Pooled female children	9.23(12/130)		P 101000	
Pooled male children	9.09(11/121)			
Pooled female children	9.23 (12/130)	0.0325 df = 1	p>0.857	

Table 4. The percentage of human African trypanosomosis seropositive residents of tsetse infested areas according to gender and age-groups in Juba area, Central Equatoria State, Sudan.

Classification of individuals	% sero-prevalence	Chi ² value	Probability, P
Pooled adults	9.05 (20/221)	1.01 df = 1	p>0.314
Pooled children	4.35 (3/69)		
Pooled adult males	9.30 (16/172)	1.371 df = 1	p>0.971
Pooled adult females	8.16 (4/49)		
Pooled adult males	9.30 (16/172)	0.793 df = 1	p>0.373
Pooled male children	2.94 (1/34)		
Pooled adult females	8.16 (4/49)	0.0 df = 1	p = 1
Pooled female children	5.71 (2/35)		
Pooled male children	2.49 (1/34)	0.0007 df = 1	p>0.979
Pooled female children	5.71 (2/35)		

Classification of individuals	% sero-prevalence	Chi ² value	Probability, P
Pooled adults	7.05 (66/936)	10 20 df - 1	n <0.00126
Pooled children	15.55 (37/275)	10.39 01 = 1	p<0.00120
De slad odult moles			
Pooled adult males	5.53 (39/705)	9.144 df = 1	p<0.00249
Pooled adult females	11.69(27/231)		1
Pooled adult males	5.53 (39/705)	7 331 df = 1	n<0.0067
Pooled male children	11.27 (23/204)	1.001 01 - 1	p <0.0007
	44.00(07/004)		
Pooled adult females	11.69(27/231)	2 339 df = 1	n>0 126
Pooled female children	8.33(14/71)	21000 41 - 1	pr 01120
Pooled male children	11 27 (23/204)		
	(1.27(23/204))	2.54 df = 1	p>0.11
Pooled temale children	8.33(14/71)	•	

Table 5. The percentage of human African trypanosomosis seropositive inhabitants of tsetse free areas according to gender and age-groups in Juba area, Central Equatoria State, Sudan.

analysis showed a significant positive correlation between the square of age of the internally-displaced female respondents (x) and their HAT sero-prevalence (y) (Y = 3.457 + 0.0002x; r + 0.9418; df =28; t = 14.077; p < 0.000001). No similar siignificant correlations could be established between age and HAT seroprvalence in any other group, irrespective of type of residence (p > 0.2 - 0.9).

Variation of human African trypanosomosis seroprevalence with vegetation type

The results shown in Table 6 reveal that the respondents examined along the riverine vegetation had a statistically higher encounter with the disease compared with those in the derived savanna woodland or the open savanna woodland.

Sero-prevalence of human African trypanosomosis based on location of respondents

The pooled sero-prevalence of the disease was relatively high in Lologo, Khor Rommalla, Tokiman, Gumbo and Kapu locations where the internally-displaced people were being settled at the time of the survey. Analysis of sero-prevalence data obtained on the displaced people showed a significant difference between settlers locations, probably highlighting their encounter with varing trypanosomosis challenge either at their abandoned original homes or enroute to these camps. Similar treatment of data, pertaining to military personnel (all adult males) who were prepetually (and are) on the move showed a relatively high sero- prevalence rate. As with the displaced people, there was also significant variation between locations. In contrast, Table 7 clearly suggests that the encounter of the diseases among residents in established villages and hamlets was relatively low with insignificant difference in sero-prevalence rates between these villages.

Relationship between human African trypanosomosis sero-prevalence and density of *G. fuscipes fuscipes*

Regression analysis (Table 8) showed no significant correlation between the density of *G. f. fuscipes* and the sero-prevalence of HAT in Southern Sudan (r + 0.698; p = 0.191; df = 8).

DISCUSSION

An active surveillance was conducted in reachable villages and camps around Juba, the Central Equatoria State capital. Under civil conflict circumstances time and efficiency are vital factors. For these reasons the card-agglutination test for trypanosomosis (CATT), is a serologic technique, highly sensitive and efficient for diagnosis of *T. b. gambiense* sleeping sickness (Magnus et al., 1978) and recommended for active surveillance to estimate the disease sero-prevelance rate was adopted (W H O, 1998). Moreover, the test is quick, easy to perform and permits the examination of hundreds of people each day (Chappuis et al., 2002, 2004).

In the present study some 2322 consenting individuals in Juba area were screened for *gambiense* HAT. Of these 257 (11.1%) were seropositive. This level of seroprevalence is almost identical to those levels regarded as epidemics in Western Equatoria (Moore et al., 1999;

Vegetation	% sero-prevalence	Chi ² value	Probability, P
Open savanna inhabitants	4.93 (17/345)	82.13 df = 2	p<0.000
Derived savanna inhabitants	7.04 (78/1108)		
Riverine vegetation inhabitants	18.64 (162/869)		
Open savanna inhabitants	4.93 (17/345	1.59 df = 1	p>0.207
Derived savanna inhabitants	7.04 (78/1108)		
Open savanna inhabitants	4.93 (17/345)	35.87 df = 1	p<0.000
Riverine vegetation inhabitants	18.64 (162/869)		
Derived savanna inhabitants	7.04 (78/1108)	60.39 df = 1	p<0.000
Riverine vegetation inhabitants	18.64 (162/869)		

 Table 6. Comparisons between human African trypanosomosis seropositive individuals in open savanna woodland, derived savanna woodland and riverine vegetation, Juba area, Central Equatoria State, Sudan.

Table 7. The perecutage of human African trypanosomosis seropositives according to locations and residence in Juba area, Central Equatoria State, Sudan.

Residence	Village	% sero-prevalence	Chi ² value	Probability, P
	Lologo	38.03 (81/213)		
	Khor Rommalla	21.36 (22/103)		
Displaced comps	Tokiman	14.81 (4/27)	02 02 df - 4	n <0.000
Displaced callips	Gumbo	7.04 (7/94)	93.92 ul = 4	μ<0.000
	Kabu	5.92 (17/287)		
	Sub-total	18.09 (131/724)		
	Rejaf East	12.5 (25/200)		
	Kit	7.69 (1/13)		
	Logo East	8.22 (6/73)		
Permanent resident	Rejaf West	12.5 (11/88)	1.698 df =5	p>0.79
	Nyori	8.62 (5/58)		
	Luri	0 (0/58)		
	Sub-total	9.8 (48/490)		
	Nyamini	15.14 (28/185)		
Mobile military camps	Koda	8.48 (15/177)		
	Serimon	1.75 (5/285)	00.00 K (
	El Jebalain	4.76 (8/168)	32.68 df = 4	p <0.000
	Rokon	7.51 (22/293)		
	Sub-total	7.04 (78/1108)		

Moore and Richer, 2001). It is suggested that the break down of the health service and the displacement of the population as a consequence of the civil conflict has aggravated the situation of sleeping sickness in the area. During the last decade there has been a similar strong resurgence of human African trypanosomosis in several countries including Angola and the Democratic Republic of Congo (Van Nieuwenhove et al., 2001; Chappuis et al., 2002). The major cause of these flare-ups is the collapse of sleeping sickness control programmes as a result of strife in these countries (Ekwanzala et al., 1996; Chappuis et al., 2002).

In the surveillance higher sero-prevalence rates were recorded in adults rather than in children. Similar results were obtained during a *T. b. gambiense* sleeping sickness surveillance in the Ivory Coast (Stanghellini and Duvallat, 1981). The high sero-prevalence of the disease in adults might be attributed to the community social behaviour and daily activities which take the adult into close and more frequent contact with the vector *G*.

Table 8. The sero-prevalence rate of human African trypanosomosis in relation to *G. fuscipes fuscipes* apparet density in Central Equatoria State, Sudan.

Location	Fly apparent density	Sero-prevalence rate	
Rejaf East	0.0	12.50 (25/200)	
Kit	3.5±0.65	07.69 (01/13)	
Logo Eat	4.0±1.15	08.22 (06/73)	
Rejaf West	1.3±0.88	12.50 (11/88)	
Nyori	5.0±1.15	08.22 (05/58)	
Luri	0.0±0.30	00.00 (0.0/58)	

Y = 2.826 + 1.62x; r + 0.696; df + 4; t + 1.681; *P*>0.191

Fly apparent density = (male +female tsetse/trap/day)

Sero-prevalence rate =(total number of seropositives divided by total examined at a location, between parentheses)

fuscipes fuscipes in watering sites. In general terms, the frequency of human/fly contact decides the disease incidence rate (Gouteux, 1985).

In the present work, the sero-prevalence rates were found to be significantly higher in adult females and children compared with that of adult men (p < 0.0005; $Chi^2 = 9$). It has been observed that there are substantial variations between men and women in the type and duration of their daily activities within the local tsetse biotopes where the disease might be transmitted, possibly resulting in profound differences in exposure between men and women and the children who, usually pursue their mothers. The adult men are mainly traders, agropastoralist and junior workers in government offices thus they have lower intensity of fly-contact compared with women. Recently, there has been considerable interest towards the influence of sex or gender on tropical diseases, sex referring to a biological characteristic while gender denotes socially constructed behaviour, expectations and roles that derive from, but may not depend on sex (Vlassoff and Manderson, 1998). It has been perceived that women had higher apparent incidence of Gambian trypanosomosis than men due to their selective activities which take them into contact with tsetse e.g. water collection, washing, firewood cutting and bathing. Children, especially girls, usually follow their mothers (Pepin et al., 2002). Dissimilarity in incidence of African trypanosomosis between genders was reported in many countreies. For instance in Fankana-Kalakitini, Kwamouth and Nioki foci of the Congo women had higher incidence of trypanosomosis than men (Henry et al., 1982; Pepin et al., 2002). Conversely, in other African countries, the incidence was higher in males in some foci. The differences in population structures of endemic villages as well as the differences in exposure to infected tsetse between men and women probably explain most of the gender variation in the incidence of HAT (Pepin and Meda, 2001).

In the present work the HAT sero-prevalence rate of the internally-displaced people and military was signi-ficantly higher than that scored for the resident population.

This might be due to the fact that the internally-displaced people and the military had probably come from highly endemic foci of sleeping sickness in Western Equatoria where they might have harboured the infection before they migrated or operated as a result of the civil unrest. The role of the population sectors who are perpetually on the move in the spread of HAT is well established in the literature and had already been discussed (Adekolu-John, 1978). In our study, the high prevalence of the disease recorded in certain locations and its absence or low prevalence in others, might be attributed to the presence of the internally-displaced people -including the government soldiers, the presence or absence of tsetse in the location and the daily activity of residents in endemic areas. Thus, the differences in the seroprevalence rates of T. b. gambiense in Juba area is in tandem with patterns of the disease in Central Africa including Uganda and the Kenya shores of lake Victoria (Hide, 1999; Van Nieuwenhove et al., 2001).

Significantly, higher prevalence of HAT was detected in residents who dwell close to or visit rivers and watercourses where *G* .*f. fuscipes* exists: in riverine galleries, thickets and farm hedges. Thus, people living or visiting these fly haunts are exposed to higher frequency of tsetse bites and hence, are at a higher risk of contracting sleeping sickness (Mulligan, 1970; Gouteux, 1985). Moreover, the war has forced the residents to flee their villages to hide in the riverine vegetation where *G. f. fuscipes* occurs.

It is well known that the *palpalis* group tsetse are more dependent on riverside vegetation (De La Rocque et al., 2001) and much less affected by human settlement. They are also better able to adapt to transformed environments as long as the vegetation cover provides them with appropriate habitats. Indeed G. f. fuscipes, a riverine tsetse, was reported to adjust itself in changing habitats (Okoth,1982). Due to their opportunistic feeding habits, the palpalis tsetse are also able to adapt from a preference for feeding on wild animals to feeding mainly on reptiles, humans and domestic animals (Cuisance et al., 1973; Jordan, 1989; Mohamed-Ahmed and Odulaja, 1997; Clausen et al., 1998). They are also more capable of adapting to peri-domestic habitats (Toure, 1974; Kuzoe et al., 1985). Adaptation to peri- domestic habitats together with the opportunistic feeding habits enable G. f. fuscipes to survive and to tolerate the adverse effect of game animals rarity (Weitz, 1963, 1970; Boyt et al., 1978; Thus, creation of peri-domestic Okoth, 1986). environments suitable for the fly might result in new HAT epidemiological systems closly linked to humans (De La Rocque et al., 1998). Indeed, due to the civil strife in southern Sudan people were settling in hamlets along the watercourses concealed in the riverine vegetation for security reasons. This may explain why the seroprevalence rate was higher in riverine vegetations rather than savanna areas or inside villages.

Correlation analysis showed no significant association

between HAT sero-prevalence rate in a location and the density of G. f. fuscipes in that location (Table 8). This finding agrees with most reports from sleeping sickness foci elsewhere in Africa where no significant relationship was demonstrated between the incidence of Gambian sleeping sickness in inhabitants and the density of species of riverine tsetse (Gouteux et al., 1993). Conversely, in the Central African Republic the rate of T. b. *aambiense* transmission was found to be significantly correlated with the density of the vector fly (Leak, 1999). The above results confirm the presence of T. b. gambiense sleeping sickness in Juba area but do not eliminate the possibility of infection with Trypanosoma brucei rohdesiense, as CATT appears to be sensitive for the former only. The Sudan lies in the interface of the geographical location of the two types of sleeping sickness (Baker et al., 1970). Considering the instability of people and their livestock due to the war there is surely a high likelihood of the spread and overlap of the two types of sleeping sicknesses. Furthermore, some strain of T. b gambiense lack the specific gene coding for CATTantigen (Penchenier et al., 2003), consequently, the level of the seropositive obtained here almost cer-tainly underestimated the true prevalence of infection. For these reasons the use of other diagnostic devices and procedures that might help to detect the true infection with both T. b. gambiense and T. b. rohdesiense are demanded.

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REFERENCES

- Adekolu-John EO (1978). The significance of migrant Fulani for human trypanosomiasis in Kainji lake areas of Nigeria. Trop. Geogr. Med., 30: 285-293.
- Baker JR, Mc Connel E, Kent DC, Hady J (1970). Human Trypanosomiasis in Ethiopia. Trans. Royal Society Trop. Med. Hygiene, 64: 523-530.
- Bloss JFE (1960). The history of sleeping sickness in the Sudan. Proceedings Royal Society Med., 53: 421-426.
- Boyt WP, Mackenzie PK, Pilson RD (1978). The relative attractiveness of donkeys, cattle, sheep and goat to Glossina morsitans morsitans Westwood and G. pallidipes Austen (Diptera: Glossinidae) in a middleveld area of Rhodesia. Bull. Entomol. Res., 68: 497-500.
- Challier A, Eyraud M, Lafaye A, Laveissiére C (1977). Amélioration du rendement du piége biconique pour glossines (Diptera, Glossinidae) par emploi un cône inférieur bleu. Cahiers ORSTOM, série Entomologie médicate et parasitologie, 15: 283-286.
- Chappuis F, Pitter A, Bovier PA, Adams K, Godinequ V, Hwang SY, Magnus E, Buscher P (2002). Field evaluation of the CATT/Trypanosoma brucei gambiense on blood-impregnated filter

- papers for diagnosis of human African trypanosomiasis in southern Sudan. Trop. Med. Int. Health, 7(11): 942-948.
- Clausen PH, Adeyemi I, Bauer B, Breloeer M, Salchow F, Staak C(1998). Host preferences of tsetse (Diptera: Glossinidae) based on blood meal identification. Med. Vet. Entomol., 12: 169-180.
- Cuisance D, Itard J, Boreham PFL (1973). Comportement des mâles steriles de Glossina tachinoides West., laches dans les conditions naturelles. Environs de Fort Lamy (Tchad)., 111. Lieux et hauteurs de repos; comportements alimentaries. Revue Elevage et de Médecine Vétérinarire des Pays Tropicaux, 26: 323-338.
- De La Rocque S, Augusseau X, Guillobez S, Michel V, De Wispelaere G, Bauer B, Cuisancw D (2001). The changing distribution of two riverine tsetse flies over 15 years in an increasingly cultivated area of Burkina Faso. B. Entomol. Res., 91: 157-166.
- De La Rocque S, Lefrancois T, Rrifenberg JM, Solano P, Kabore I, Bengaly Z, Augusseau X, Cuisance D (1998). PCR analysis and spatial repartition of trypanosomes infecting tsetse flies in Sideradougou area (Burkina Faso). Annal. New York Acad. Sci., 849: 32-38.
- Ekwanzala M, Pepin J, Khonde N, Molisho S, Bruneel H, De Wals P (1996). In the heart of darkness: sleeping sickness in Zaire. Lancet, 348: 1427-1430.
- Ford J (1971). The Role of Trypanosomiasis in African Ecology, Oxford, Clarendon Press.
- Gouteux JP (1985). Ecologie des Glossines en secteur pré-forestiére de Côte Ivoire Relation avec la trypanosomiases humaine et possibilités de lutte. Annales de Parasitoloie Humaine et Comparée, 60: 329-347.
- Gouteux JP, Kounda JC, Gboumi L, Noutoua D, Amico F, Bailly C, Roungou JB (1993). Man-fly contact in the Gambian trypanosomiasis focus of Nola-Bilolo (Cenral Africa Republic). Trop. Med. Parasitol., 44: 213-218.
- Hargrove JW, Langley PA (1990). Sterilising tsetse (Diptera: Glossinidae) in the field: a successful trial., Bull. Entomol. Res., 80: 397-403.
- Henry MC, Ruppo JFI, Bruneel H (1982). Distribution de infection Par T. b. gambiense dans une population du Bandundu en République du Zaire. Annales de la Société Belge de Médecine Tropicale, 62: 301-313.
- Hide G (1999). History of sleeping sickness in East Africa. Clin. Microbiol. Rev., 12(1): 112-125.
- Hunt AR, Bloss JFE (1945). Tsetse fly control and sleeping sickness in the Sudan. Trans. Royal Society Trop. Med. Hygiene, 39: 43-58.
- Hutchinson MP (1975). Assignment report: trypanosomiasis in southern Sudan. 29 April1974 - 31 March 1975. WHO, EM/PD/5 (or EM/SUD/MPD/005/FR (UNHCR) (2301)) May 1975.
- Jordan AM (1989). Importance of land use on the incidence of Trypanosoma brucei gambiense sleeping sickness. Annales de la Société Belge de Médecine Tropicale, 69(1): 254.
- Kuzoe FAS (1991). Perspectives in research on and control of African trypanosomiasis. Ann. Trop. Med. Parasitol., 85(1): 33-41.
- Kuzoe FAS (1993). Current situation of African trypanosomiasis. Acta Tropica, 54: 153-162.
- Kuzoe FAS, Baldry DAT, Van Der Vloedt A, Cullens JR (1985). Observation of an apparent population extension of Glossina tachinoides Westwood in southern Ivory Coast. Insect Sci. its Appl., 6: 55-58.
- Leak SGA (1999). Tsetse Biology and Ecology: Their role in the epidemiology and control of trypanosomosis. ILRI, Nairobi, Kenya. CAB Publishing.
- Magnus E, Vervoort T, Van meirvenne N (1978). A card- agglutination test with stained trypanosomes (CATT) for serological diagnosis of T.
 b. gambiense trypanosomiasis. Annales de la societe belge de Medecine Tropical., 58: 169-196.
- Mohamed-Ahmed MM, Odulaja A (1997). Diel activity patterns and host preferences of Glossina fuscipes fuscipes along the shores of Lake Victoria, Kenya. Bull. Entomol. Res., 87: 179-186.
- Moore A, Richer M (2001). Re-emergence of epidemic sleeping sickness in southern Sudan. Trop. Med. Int. Health, 6(5): 342-348. Moore A, Richer M, Enrile M, Losio E, Roberts J, Levy D (1999).
- Resurgence of sleeping sickness in Tambura county, Sudan. Am. J. Trop. Med. Hygiene, 61: 315-318.

- Mulligan HW (1970). The African Trypanosomiasis, London, George Allen and Unwin. Odulaja A, Mohamed-Ahmed MM (1997), Estimation of the efficiency of the Biconical trap for Glossina fuscipes fuscipes along the Lake Victoria shore, Kenya., Entomologia Experimentalis et Applicata, 82: 19-24.
- Okoth JO (1982). Further observations on the composition of Glossina population at Lugala, South Busoga District, Uganda. East Afr. Med. J., 59: 582-584.
- Okoth JO (1986). Peridomestic breeding sites of Glossina fuscipes fuscipes Newst. In Busoga,Uganda, and epidemiological implications for trypanosomiasis. Acta Tropica, 43: 283-286.
- Penchenier L, Grébaut P, Njokou F, Eboo Eyenga V, Büscher P (2003). Evaluation of LATEX /T.b. gambiense for mass screening of Trypanosoma brucie gambiense sleeping sickness in Central Africa. Acta Tropica, 85(1): 31-37.
- Pepin J, Meda H (2001). The epidemiology and control of African trypanosomiasis. Adv. Parasitol., 40: 71-132.
- Pepin J, Mpia B, Iloasebe M (2002).Trypanosoma brucie gambiense African trypanosomiasis: differences between men and women in severity of disease and response to treatment. Trans. Royal society Trop. Med. Hygiene, 96: 421-426.
- Snow WF (1983). Assignment Report: Tsetse distribution and ecology in relation to sleeping sickness in Southern Sudan. May- June 1982, WHO, EM/PD/5: EM/SUD/MPD/005/ FR (UNHCR) (2301).

- Snow WF (1984). Assignment Report: Further observation on tsetse distribution, ecology and control in relation to sleeping sickness in southern Sudan., 12 February- 13 June 1983, WHO.
- Stanghellini A, Duvallat G (1981). The epidemiology of human trypanosomiasis due to Trypanosoma gambiense in a focus of the Ivory Coast. 1. The distribution of the disease in the population. Z. Tropenmed. Parasit., 32(3): 141-144.
- Toure SM (1974). Note sure quelques particultarites dans habital de Glossina palpalis gambiensis (Diptera : Glossinidae) observées au Sénégal. Revue Elevage et de Médecine Vétérinarire des Pays Tropicaux, 27: 81-94.
- Van Nieuwenhove S, Betu-Ku-Mesu VK, Diabakana PM, Declercq J, Bilenge C MM (2001). Sleeping sickness resurgence in the DRC: the past decade. Trop. Med. Int. Health, 6(5): 335-341.
- Vlassoff C, Manderson L. (1998). Incorporating gender in the anthropology of infectious diseases. Trop. Med. Int. Health, 3: 1011-1019.
- Weitz B (1963). The feeding habits of Glossina. Bull. World Health Organ., 28: 711-729.
- Weitz B (1970). Hosts of Glossina.: The African Trypanosomiases,
- London, George Allen and Unwin Ltd., pp: 416-423
- World Health Organization (1998). Cnotrol and Surveillance of African Trypanosomiasis. WHO Technical Report Series 881, WHO, Geneva.