

Full Length Research Paper

## Binding pattern of ferrocypen upon interaction with cetyltrimethyl ammonium bromide in the presence of urea

Olaseni Segun Esan<sup>1\*</sup>, Christopher Olumuyiwa Aboluwoye<sup>1</sup>, Nurudeen Abiola Oladoja<sup>1</sup>; Jide Ige<sup>2</sup>, Olanrewaju Owoyomi<sup>2</sup> and Grace Olufunmilayo OGUNLUSI<sup>2</sup>

<sup>1</sup>Department of Chemistry and Industrial Chemistry, Adekunle Ajasin University, Akungba Akoko Nigeria.

<sup>2</sup>Department of Chemistry, Obafemi Awolowo University Ile-Ife Nigeria.

Accepted 04 April, 2017

Cetyltrimethylammonium bromide (CTABr) have been titrated against Ferrocypen (Dicyano-bis-(1, 10-phenanthroline) iron II complex) in aqueous solution as a function of urea concentration which gave a sigmoidal binding isotherm. A simple method was introduced for resolution and characterization of binding set on the basis of binding capacity concept to give hill coefficient > 1.0. Hill coefficient greater than one are the experimental hall mark of cooperativity in which initial binding events render subsequent binding events more favorable. The degree of cooperativity is sensitive to the concentration of urea. The results are interpreted in terms of dielectric constant and decrease in the hydrophobic interaction between the complex (ferrocypen) and the surfactant monomers.

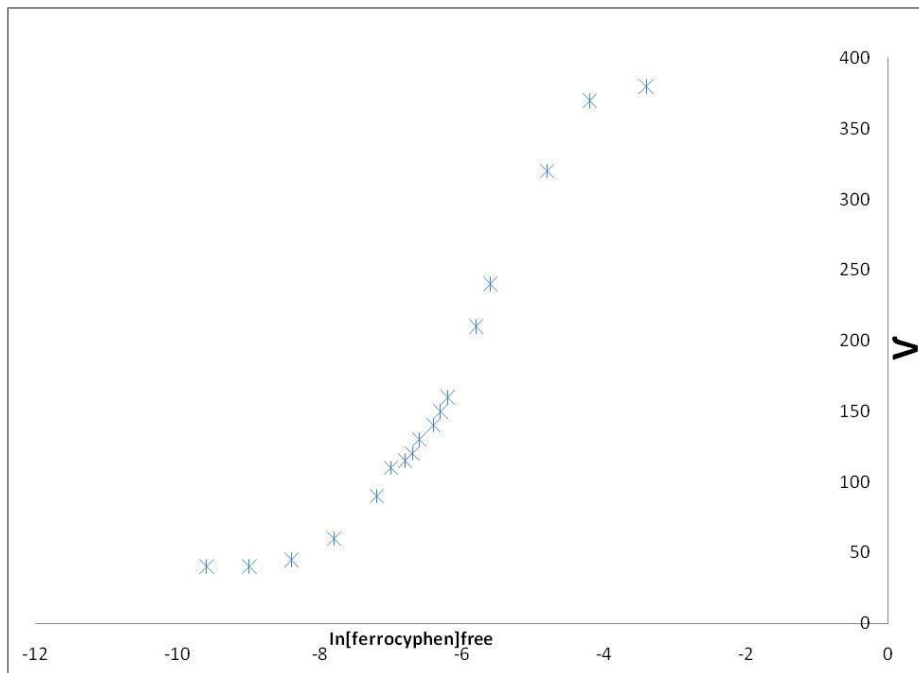
**Key word:** Binding isotherm, surfactant, cooperativity, hydrophobic ferrocypen, binding capacity.

### INTRODUCTION

Allosteric interaction between recognition sites is a ubiquitous regulatory mechanism in biological macromolecules, including enzyme (Fersht, 1985), receptor (Falke and Koshland, 1987) and ribosome's (Nierhaus et al., 1988). Micelles are thought to mimic the active site in enzyme (Ige et al., 2007) and when the active and effect-tors site are equivalent, the allostery is termed cooperativity for which enzymes is the typical examples (Russell et al., 1990; Monod et al. 1963; Eigen and Nobel, 1967; Rebek, 1984). The ability of micelles to alter the physico-chemical properties of solutes has been extensively used in both equilibrium (Rudoif et al., 1987; Hinze, 1979) and kinetic determination and different separation techniques (Okada, 1997; Corstjens, 1995). Cooperative binding of amphiphilic substances such as surfactants and dyes to macromolecules is a very important phenomenon used to study colloidal properties and initial denaturation stage

of the macromolecules (Kiyofumi, 2007). Urea is known to moderate the aggregation of surfactant monomer (Ana et al., 2005; Shashank et al., 2001). The neutral bulky ferrocypen with significant hydrophobic character and Cetyltrimethyl ammonium bromide (CTABr) were specially selected for their overall varying hydrophobicity (Ige et al., 2007). For biological system where similar hydrophobic interactions are known, the formation of a hydrophobic core is the driving force for protein folding. It is possible by choice of the appropriate proteins to ensure only hydrophobic interaction (Chad and Marcey, 2004). This type of selectivity in hydrophobic binding has a more direct correlation to micelle-substrate binding system. It is therefore of interest to see how the moderation of protein folding via hydrophobic and hydrogen bond breaking by urea is mimic in a simple surfactant system. In the present study, the interaction of Ferrocypen with Cetyltrimethylammonium bromide in the presence of urea has been studied using spectrophotometry techniques (Housaindokht, 2001). Replotting of the binding isotherm gives Hill coefficient (greater than) > 1 which is the hallmark of cooperativity.

\*Corresponding author. E-mail: [lincolnolaseni@yahoo.com](mailto:lincolnolaseni@yahoo.com). Tel: +2347030077310



**Figure 1.** Binding isotherm of ferrocypen interaction with CTABr in the presence of Urea at 25°C.

**EXPERIMENTAL**

**Materials**

Cetyltrimethylammonium bromide (CTABr) was used as supplied without further purification. The degree of purity was ascertained by determining the critical micelle concentration (CMC) in aqueous medium at 25°C. The value  $9.1 \times 10^{-4} \text{ moldm}^{-3}$  was obtained which is in good agreement with the reported literature value (Peterson and Marzzacco, 2007).

1,10-phenanthroline, ferrous ammonium sulphate, urea and potassium cyanide were analytical grade (Sigma). Ferrocypen was synthesized as reported in the literature (Alfred, 1960). The water used was glass distilled.

**METHODS**

The binding studies were carried out at a fixed temperature of  $25.0 \pm 0.1^\circ\text{C}$ . A wavelength scans of the complex in urea/ water/surface-tant medium show no change in  $\lambda_{\text{max}}$  under the present experimental condition. Noticeable increase in absorbance was observed at fixed ferrocypen and urea concentration with varying surfactant concentration. Fraction of the ferrocypen bound was calculated from these changes in absorbance. All binding studies were carried out at surfactant concentration below the critical micelle concentration. The change in absorbance of the ferrocypen was monitored using an - Helios Pye-unicam double beam spectrophotometer fitted with a thermostable cell compartment. The absorbance was taken at 560 nm (the wavelength of maximum absorption) of Ferrocypen in all the runs. The results were analyzed using binding capacity concept.

Binding capacity is the homotropic second derivatives of the binding potential with respect to the chemical potential of the ligand

( $\mu_s$ ) and provides a measure of steepness of the binding isotherm (Bordbar et al., 2004).

It represents the change in the number of moles of ligand per mole of macromolecule ( $v$ ) that accompanied a change in the chemical potential of that ligand.

**RESULTS AND DISCUSSION**

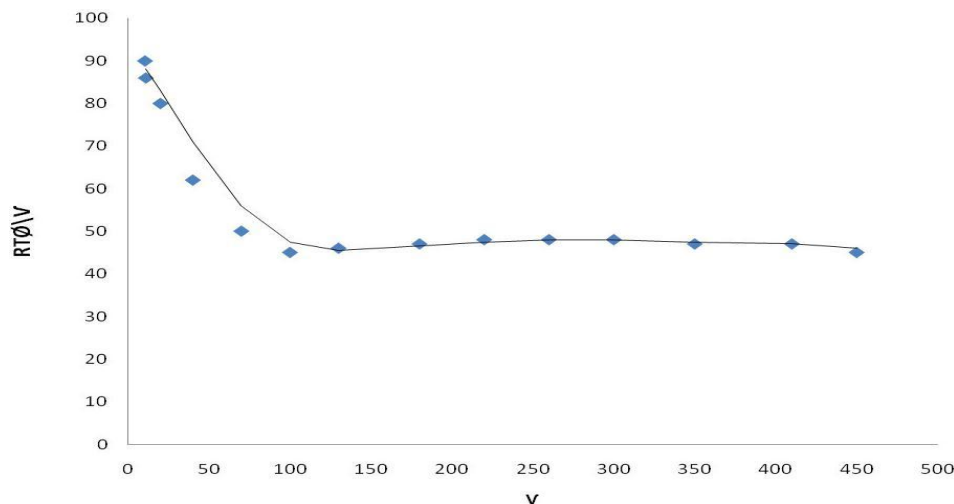
Figure 1 represent binding isotherm for a typical interaction of Ferrocypen with Cetyltrimethyl ammonium bromide in the presence Urea and shows the variation of  $v$  (average number of ferrocypen bound to Cetyltrimethyl ammonium bromide molecule) versus  $\ln[\text{ferrocypen}]_{\text{free}}$  at specified experimental condition. The sigmoidal nature of this curve characterized cooperative binding (Cera et al., 1988).

The average number of ferrocypen molecules bound to surfactant molecules has been calculated as,

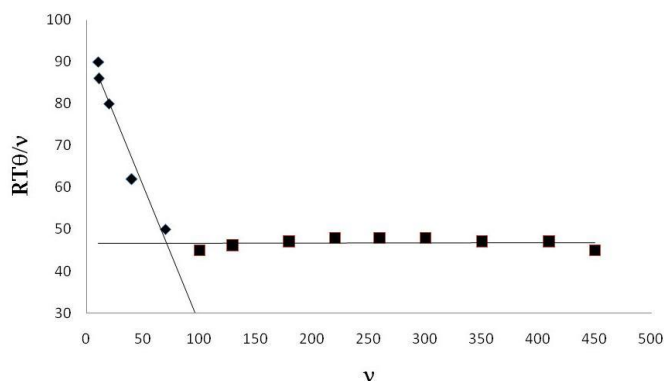
$$v = \frac{[\text{Ferrocypen}]_{\text{T}} - [\text{ferrocypen}]_{\text{f}}}{[\text{Surfactant}]_{\text{T}}} \tag{1}$$

Where  $[\text{Surfactant}]_{\text{T}}$  is the total concentration of surfactant.  $[\text{Ferrocypen}]_{\text{T}}$  and  $[\text{ferrocypen}]_{\text{f}}$  are the total and free concentration of ferrocypen respectively. By considering the ideal behavior, binding capacity ( $\theta$ ) equal to

$$\theta = (\partial v / \partial \mu)_{\text{T,P},\mu_j} = (\partial v / RT \partial \ln[L]_{\text{F}})_{\text{T,P},\mu_j} \tag{2}$$



**Figure 2.** The plot of  $RT\theta/v$  versus  $v$  for binding of ferrocypen with CTABr in the presence of urea at  $25^{\circ}\text{C}$ .



**Figure 3.** The plot of  $RT\theta/v$  versus  $v$  for the interaction of ferrocypen with CTABr at  $25^{\circ}\text{C}$  in the presence of urea.

Where  $R$ ,  $T$  and  $[L]_f$  are gas constant, absolute temperature, and free concentration of Ferrocypen<sup>24</sup>. However binding capacity can be estimated by calculating the steepness of the binding isotherm. This concept is directly related to the type and extent of cooperativity as expected by Hill coefficient,  $n_{H_i}$  (Reza and Ramin, 2007). The value of  $\theta$  at any  $v$  can be determined by calculating the slope of the binding isotherm. The relationship between binding capacity  $\theta$ , and Hill coefficient  $n_{H_i}$  can be obtain by assuming a particular definition for  $n_{H_i}$ .

The Hill coefficient is defined as the slope of Hill graph,

$$n_H = \frac{d \ln \left( \frac{\theta}{1-\theta} \right)}{d \ln [L]_f} = \frac{\left( \frac{1}{\theta(1-\theta)} \right) d\theta}{d \ln [L]_f} \quad (3)$$

Where  $y$  is the fractional saturation of the macromolecule

by the ligand which is defined as follows.

$$y = v/g \quad (4)$$

From the definition of binding capacity, (i.e equation 1) the following equation can be written

$$n_H = RT\theta/gy(1-y) \quad (5)$$

$$\theta = n_{H_i} v (1-y)/RT \quad (6)$$

Equation (6) can be rearranged to the following form

$$RT\theta/v = n_H - n_{H_i} v/g \quad (7)$$

This equation suggests that the plot of  $RT\theta/v_i$  versus  $v_i$  for a system with  $g$  identical and independent sets, should be linear. The slope and the Y and X-Intercepts are equal to  $-n_{H_i}/g$ ,  $n_H$  and  $g$  respectively. Where  $g_i$  and  $n_{H_i}$  are the number of binding sites and Hill coefficient for  $i$ th binding set respectively.

Figure 2 shows the variation of  $RT\theta$  vs  $v$  for binding of Ferrocypen to Cethyltrimethyl ammonium bromide. For a system with relatively high difference in binding affinity of sets, every set behave independently, and so in which it can be assume  $i$ th the  $i$ th binding set has not been occupied until the full occupation of  $(i-1)$ th binding set has been occurred, for such system the curve of  $RT\theta/v$  vs  $v$  can be divided into  $N$ -consecutive linear part, corresponding to  $N$  binding sets. For evaluation of this assumption, the typical plot of  $RT\theta/v_i$  VS  $v_i$  have been constructed for binding of Ferrocypen to Cethyltrimethyl ammonium bromide at different experimental condition (Figure 3). Knowing  $n_{H_i}$  and  $g_i$ , the Hill plot were constructed for the estimation of Hill binding constant  $K_i$  by plotting  $\ln(v_i/g-v_i)$  against  $\ln[\text{Ferrocypen}]_{\text{free/mol dm}^3}$ .

**Table 1.** The collective values of Hill parameters for interaction of ferrocypen with CTABr at 25°C

[Urea]M	n <sub>H1</sub>	g <sub>1</sub>	K <sub>1</sub> (M <sup>-1</sup> )x10 <sup>4</sup>	ΔG <sup>1</sup> <sub>b,v</sub> x 10 <sup>3</sup>	n <sub>H2</sub>	g <sub>2</sub>	K <sub>2</sub> (M <sup>-1</sup> )x10 <sup>4</sup>	ΔG <sup>2</sup> <sub>b,v</sub> x 10 <sup>3</sup>
0	16.73	181.07	11.50	-9.14	3.22	417.03	12.17	-18.12
4.0	41.31	203.11	10.17	-8.04	38.31	420.13	11.51	-22.14
5.0	51.02	213.01	9.88	-23.50	45.71	422.05	15.01	-35.40
6.0	20.78	114.16	27.88	-7.13	16.56	98.45	37.33	-10.23
8.0	13.01	93.67	34.98	-7.11	10.25	64.92	52.71	-6.01

Figure 3 shows the Hill plots for interaction of Ferrocypen with Cethyltrimethyl ammonium bromide in the present of urea at 25°C presence of urea. The intrinsic Gibbs free energy of binding per mole of ferrocypen for *i*th binding set, ΔG<sup>(i)</sup><sub>b,v</sub>, can be calculated by the following equation (Bordbar et al., 1997)

$$\Delta G_{b,v}^{(i)} = -RTn_{Hi}\ln K_i + RT(1-n_{Hi})\ln[\text{Ferrocypen}]_F \quad (8)$$

## Conclusion

There are at least two binding set in this system. The process shows positive cooperativity in both binding sets (n<sub>Hi</sub>> 1) for all the studied condition (Table 1) . At low Urea concentration, local dielectric constant increases (Marilyn and Alfred, 1967).

It is suggested in the present work that this will necessary accompanied by enhancement of pre-micelle formations and a subsequent increase in n<sub>Hi</sub> and g<sub>i</sub> as well as decrease in binding constant. Above a critical concentration of 6.0 M of Urea, dielectric constant has no predominant effect (reduced). Increase in the concentration of urea clearly diminished the hydrophobic interaction between Ferrocypen and the surfactant monomers by aggregating via hydrogen bonding to itself and water molecule creating cavities of various sizes where some Ferrocypen molecule are trapped and increasing the solubility of monomers, therefore delaying the onset for pre-micelle formation leading to decrease in n<sub>Hi</sub> and g<sub>i</sub> value but increase in binding constant. Overall, the evidence presented supports the direct interaction mechanism of urea and further demonstrate how the properties of surfactant aggregate can be moderated with the use of an inert additive such as urea.

## REFERENCES

- Alfred AS (1960). Mixed Ligand complexes of Iron II and III with Cyanide and Aromatic Di-imines. *J. Am. Chem. Soc.* 82: 3000-3005.
- Ana HC, Kerstin N, Kurt BD, Lennart N (2005). Effect of urea on peptide contamination in water molecules dynamics and experimental characterization. *Biophysical J.*, 89: 842-857.
- Bordbar AK, Saadati Z, Sohrabi N (2004). Analysis of Ligand binding process using binding capacity concept. *Acta. Biochimica. Polonica*, 5: 963-970.
- Bordbar AK, Saboury AA, Housaindokht MR, Moosavi-Movahedi AA (1997). Three binding set Analysis of alpha – lactalbumin by interaction of Tetradecyltrimethyl Ammonium Bromide. *J. Coll. Int. Sci.*, 192: 415-425.
- Cera E (1988). Binding capacity: Cooperativity and Buffering in Biopolymeric. *S. Gill, J. Wyman, Proc. Natl. Acad. Sci. USA*, 85: 449-452.
- Chad DT, Marcey LW (2004). Comparizon of C – H ..... and Hydrophobic interaction in a β - Hairpin peptide: Impact on stability and specificity. *J. Am. Chem. Soc.* 126: 2028-2034.
- Corstjens H, Billiet HAH, Frank J, Luyben KCA, *J. Chromatogr, A*, 1995, 715, 1-11.
- Eigen M (1967). Kinetics of reaction control and information transfer in enzymes and nucleic acid. *Nobel Symp.*, 5: 333-340.
- Falke JJ, Koshland DE (1987). Evidence that ligand Binding and Covalent Adaptation Drive a Two-State equilibrium in the Aspartate Receptor signaling complex. *J. Sci.*, 237: 1596-1599.
- Fersht A (1985). *Enzyme structure and mechanism*(2<sup>nd</sup> End);, Freeman, New York, pp 39-41.
- Hinze WL (1979). In use of surfactant and micellar system in Analytical Chemistry(Mittal, K.L Ed);, New York,1: 79-127.
- Housaindokht MR, Chamani J, Saboury AA, Moosavi – movahedi AA, Bahrololoom M (2001). A Microcalorimetry and binding study on interaction of dodecyltrimethyl ammonium bromide with Wigeon Hemoglobin. *Bull Korcan. Chem. Soc.*22: 145 -148.
- Ige JW, Olaseni ES, Owoyomi O, Ogunlusi GO, Soriyan OO (2007). Binding of Ferrocypen by SDS, CTAB and Triton X- 100 in water – Ethanol Co-solvent. *Ife J. sci.*, 9: 137-144.
- Kiyofumi M (2007). Thermodynamics and Kinetic aspects of self – association. *Dyes and pigments* 53: 31-43.
- Marilyn EF, Alfred H (1967). The Hydrophobic Bond in Micellar system. Effect of various Additives on the stability of micelles of sodium Dodecylsulphate and of n-Dodecyl trimethylammorium Bromide. *J. Phys. Chem.*, 71: 3320-3330.
- Nierhaus KH, Hausner PP, Geigenmuller U, (1988). The Allosteric three – site model for the ribosomal elongation cycle; Features and Future. *J. Biol. Chem.* 263: 103-109.
- Okada T (1997). Peptide separation in normal – phase liquid Chromatography study of selectivity mobile phase effect on various Columns. *J. Chromatogr., A*, 780: 343-360.
- Peterson B, Marzzacco CJ (2007). *The chemical Educator*, 12: 80-84.
- Reza H, Ramin M, Amir AM (2007). Interaction of Diclofenac with Bovine Serum Albumin Investigated by Diclofenac – Selective Electrode. *Acta. Chim. Slov.*, 54: 126-130.
- Rudolf S, Ewa M, Felix M (1987). . Acid – Base titrations of barbiturates in aqueous micellar media. *Anal Chim Acta.* 329: 4.
- Russell CP, Jeffrey SS, Christopher TS, Kumaravel G, Fu LT (1990). Cooper active Binding by Aggregated Mono – 6 – (alkylamino) – β – Cyclodextrins. *J. Am. Chem. Soc.*, 112: 3860-3863.
- Shashank D, Jagdish AL (2001). Interaction of bromine serum albumin with anionic surfactants. *Phys. Chem. Chem. Phys.* 3: 4533-4591.