

Full Length Research Paper

# Genetic diversity of *Uapaca kirkiana* Muel. Arg. populations as revealed by amplified fragment length polymorphisms (AFLPs)

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*Uapaca kirkiana* is a priority fruit tree species for domestication in miombo woodlands of Southern Africa. Natural populations of *U. kirkiana* are declining through out the woodlands due to deforestation, forest fragmentation and wildfires. Knowledge of population structure and genetic diversity is prerequisite for development of conservation strategies. Amplified fragment length polymorphisms (AFLP) were used to assess the genetic diversity of eight populations from three geographical regions of Malawi. AFLP markers revealed moderate differentiation ( $G_{ST} = 0.079$ ) among the populations collected from the three regions, however, there were no significant genetic variations among the regional collections. Analyses of molecular variance (AMOVA) found very high variation (92%) among individuals within populations and 6.8% among populations. The variations between populations indicate that populations can not be considered a single panmictic unit. Analyses of genetic similarity based on unweighted pair group method of arithmetic averages (UPGMA) suggested that the 8 populations fall into three clusters with 5 populations in one cluster, two in another and the lake shore population of Chesamu in its own cluster. Based on results presented it would be cost effective to sample a small number of populations represented by a large number of individuals for germplasm conservation purposes. In view of the wide distribution of *U. kirkiana* in the miombo woodlands in Southern Africa there is need for a more intensive genetic study to include populations growing in different countries to produce a wider picture of levels of distribution of genetic diversity of the species.

**Key words:** AFLP, conservation, dioecious, domestication, genetic diversity, miombo, *Uapaca kirkiana*, UPGMA.

## INTRODUCTION

*Uapaca kirkiana* Muel. Arg is a dioecious plant that belongs to family Euphorbiaceae. The genus *Uapaca* consists of around 60 species of which 49 are restricted to tropical Africa whilst the rest occur only in Madagascar (Radcliffe-Smith, 1988). The genus is stable, apparently devoid of polyploidy, with a chromosome number of  $2n=26$ . The range of *U. kirkiana* includes Angola,

Burundi, southern Democratic Republic of Congo, Malawi, Mozambique, Tanzania, Zambia and Zimbabwe (Ngulube et al., 1995). Ethnobotanical studies within the miombo ecological zone in Malawi (Maghembe and Seyani, 1992; Maghembe et al., 1994), Tanzania (Karachi et al., 1991) and Zambia (Kwesiga and Chisumpa, 1992) identified *U. kirkiana* as a priority indigenous fruit tree species for conservation and domestication among 50 species of indigenous trees bearing edible fruits. Fruits of *U. kirkiana* have high nutritional value and they can be eaten raw, made into jam and sweetmeats, or used to produce a refreshing

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**Table 1.** Eight *Uapaca kirkiana* populations collected in Malawi with climatic data for localities.

Population and code	Land tenure	Region	Latitude (°S)	Longitude (°E)	Altitude, category (m)	Rainfall (mm)
Chesamu (8)	customary	Northern	11 <sup>U</sup> 49	34 <sup>U</sup> 10'	1118 (medium)	1500
Chimaliro (6)	forest reserve	Central	12 <sup>U</sup> 28'	33 <sup>U</sup> 29'	1338 (high)	1200
Chimpeni (2)	private	Southern	15 <sup>U</sup> 22	35 <sup>U</sup> 24'	850 (low)	1300
Chongoni (4)	forest reserve	Central	14 <sup>U</sup> 19	34 <sup>U</sup> 15'	1500 (high)	1100
Dzalanyama (5)	forest reserve	Central	14 <sup>U</sup> 17'	33 <sup>U</sup> 31'	1200 (medium)	1150
Likhubula (1)	foest reserve	Southern	15 <sup>U</sup> 06	35 <sup>U</sup> 30	1260 (high)	1000
Perekezi (7)	forest reserve	Northern	12 <sup>U</sup> 29'	33 <sup>U</sup> 29'	1300 (high)	1300
Tsamba (3)	customary	Southern	15 <sup>U</sup> 49'	34 <sup>U</sup> 35'	890 (low)	850

drink and a variety of wines. The wood of *U. kirkiana* is a good source of firewood, poles for buildings, domestic utensils, furniture and carvings. In recent years, the development of cottage wine industries in Malawi and Zambia has opened an attractive market for the fruit. Large quantities are now required to sustain these industries as well as to meet the growing demand for whole fruits in rural and city markets (Ngulube et al., 1997). Despite the benefits *U. kirkiana* provide the tree species is disappearing from its natural stands due to clearing of natural forests for fuel wood, agriculture, settlement and other development schemes. Already the species has completely disappeared in some parts of Southern and Central Malawi (Ngulube and Kananji, 1989).

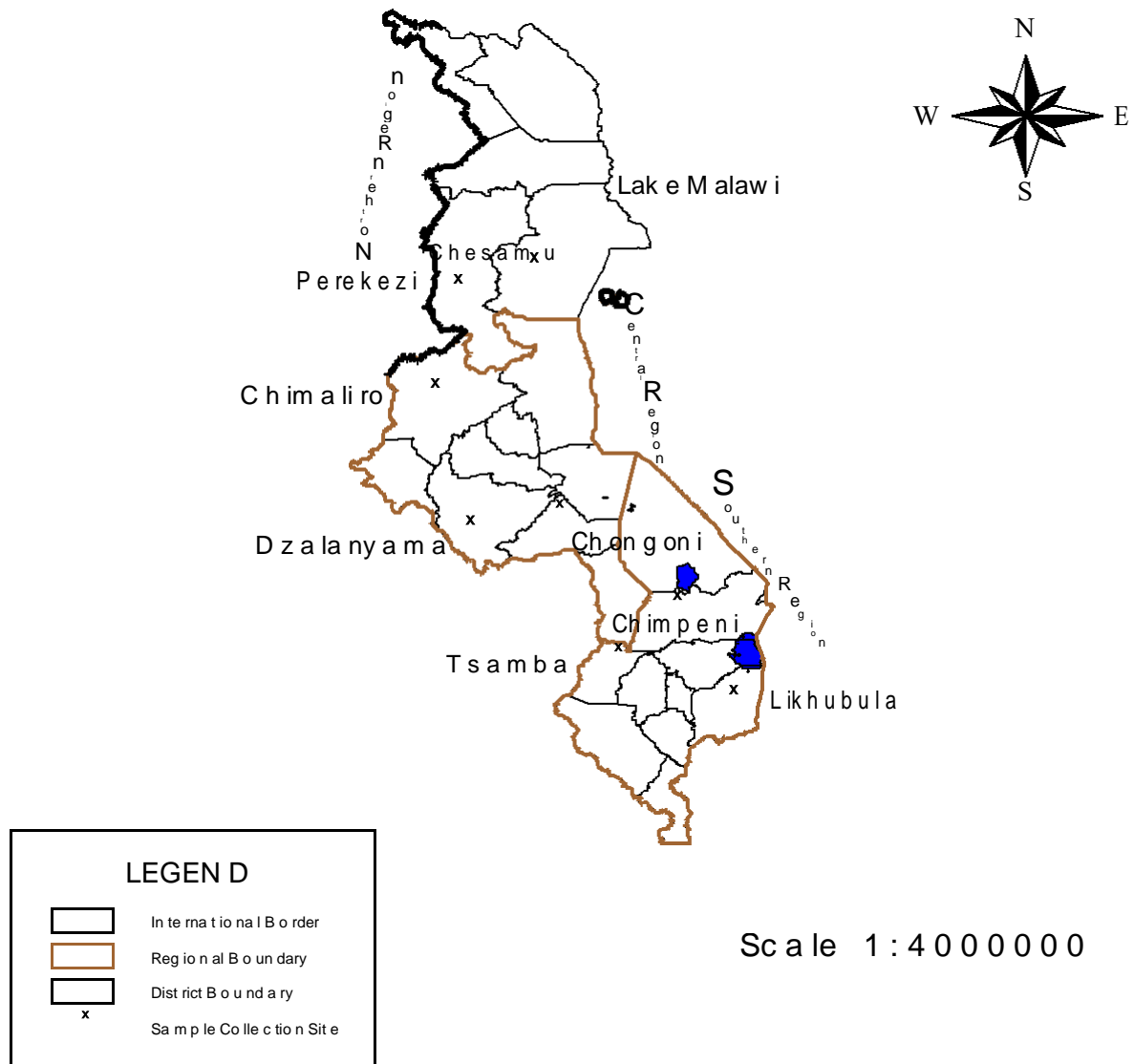
Forest reserves and national parks are under threat as some conservation areas have been degazetted in favour of agricultural activities and settlements to accommodate growing populations. A drive towards domestication of *U. kirkiana* in Southern Africa is a feasible option for sustainable conservation and management of the species. The conservation of genetic diversity both within and among natural populations is a fundamental goal of conservation biology (Hamrick et al., 1991). To manage germplasm resources effectively requires knowledge of amount and distribution of genetic diversity present in natural populations. A sustainable domestication strategy requires detailed information on the genetic structure of populations. To our knowledge, this is currently not available for *U. kirkiana*. Studies that have been done so far have concentrated on morphological variation and are, therefore, not adequate to characterise *U. kirkiana* populations existing in Malawi for selection and conservation. Large morphological variation has been reported (Mwamba, 1995; Ngulube, et al., 1997; Mwase et al., in preparation) among trees, fruits, seeds and seedlings of *U. kirkiana* from different populations. *U. kirkiana* is widely distributed across the agroecological zones of Malawi such that part of the variation observed may be attributed to differences in environmental and edaphic factors.

Molecular genetic markers provide a relatively unbiased method of quantifying genetic diversity. Molecular genetics provide fast and effective tools to biologists and geneticists to reveal diversity and variability among plants, animals and microorganisms at DNA level. Information on molecular diversity of *U. kirkiana* is limited and data on genetic variation at the DNA level of Malawian populations is lacking. Several molecular markers are available for studying genetic diversity in plants. Amplified Fragment Length Polymorphisms (AFLPs) (Vos et al., 1995) based on the polymerase chain reaction (PCR) are spread all over the genomes and are hypervariable. Use of AFLPs is advantageous in generation of large number of markers spanning the whole genome without prior knowledge of sequence of the genome. AFLPs, however have disadvantage of being less informative because of their dominance and bi-allelic nature (Heckenbecker et al., 2003). AFLPs allow relatively quick marker development, which is often important in the assessment and conservation of genetic diversity (Marghali et al., 2005). Due to their higher multiplex ratio, discriminatory power and reproducibility among different laboratories, AFLPs have been used successfully in plant population genetic studies (Muluvi et al., 1999; Schmidt and Jensen, 2000). The primary objectives of the study were to detect and measure genetic variation in natural populations of *U. kirkiana* in Malawi and examine the usefulness of AFLP marker technology in this species.

## MATERIALS AND METHODS

### Collection of plant material

A total of 96 samples were collected from 8 localities with 12 individuals representing a population (Table 1) along the geographical range of natural distribution of *U. kirkiana* in Malawi (Figure 1). Selected populations were heterogeneous for forest land tenure, habitat type and elevation. Collections were made from three land tenure categories namely public land in forest reserves, customary and leasehold land. About 5 g of young leaves were harvested from young leaves in the field and preserved in silica gel



**Figure 1.** Map of Malawi showing location of 8 *Uapaca kirkiana* populations sampled from northern, central and southern regions.

in 50 ml vials. Young leaves were sampled with minimum distance between sampled trees set as 2 m to reduce the chance of sampling from sprouts from similar individuals.

#### DNA extraction

Total genomic DNA extraction was from 200 mg leaf sample following the protocol of Patterson et al. (1993) with minor modifications. Following precipitation of impurities and RNase digestion isolated DNA was resuspended in 200 µl Tris-EDTA buffer and stored at -20°C until further analysis. DNA quality was checked by ethidium bromide staining on a 1% agarose gel and concentration was estimated by visual assessment relative to 1 Kb DNA (Gibco Life Technologies) ladder of different known concentrations (Sambrook et al., 1989).

#### AFLP analysis

AFLP reactions were performed on individual DNA samples from the different natural populations. AFLP analysis was carried out according to the protocol of Vos et al. (1995) using restriction enzymes *EcoRI* and *MseI*. Two replicates of PCR reactions from independent DNA extractions were performed to determine the accuracy of the analysis. Initially, 64 primer pairs originated by the combination of 8 *EcoRI* and 8 *MseI* primers were tested in ten *U. kirkiana* from 10 populations. From the primer screening exercise, six primer pairs (Table 2) were selected based on clarity and reproducibility of electrophoretic patterns and applied to all the 8 populations. Five microlitres of the 1:10 diluted pre-amplification was used as template for selective amplification using primers with three and four selective nucleotides at the 3' end. Two replicates of the PCR reactions from independent DNA extractions were performed to determine the accuracy of the analysis. AFLPs were

**Table 2.** Level of polymorphism and fingerprinting of AFLP markers in *Uapaca kirkiana*.

Primer name	Primer combination	Total bands	Polymorphic bands	Polymorphism rate (%)
E33/M59	<i>EcoR1</i> -AAG/ <i>Mse1</i> -CTA	92	10	10.9
E33+C/ M47	<i>EcoR1</i> -AAGC/ <i>Mse1</i> -CAA	95	13	13.7
E35/M47	<i>EcoR1</i> -ACA/ <i>Mse1</i> -CAA	107	14	7.6
E36/M62	<i>EcoR1</i> -ACC/ <i>Mse1</i> -CTT	106	16	15.1
E40+A/M60	<i>EcoR1</i> -AGCA/ <i>Mse1</i> -CTC	104	25	24.0
E41+A/M59	<i>EcoR1</i> -AGGA/ <i>Mse1</i> -CTA	108	32	3.4
	<b>Total</b>	612	110	18.0
	<b>Mean</b>	102	18.3	5.6

**Table 3.** AMOVA for 96 individuals of *Uapaca kirkiana* from eight populations in Malawi using 110 AFLP markers. Statistics include sums of squares, mean squares, variance component estimates, percentage of total variance and the probability (p) of obtaining a more extreme component estimate by chance alone (estimated from 1000 sampling realizations).

Source of variation	d.f.	SS	MS	Variance component	Total (%)	P value
Among regions	2	68.872	34.436	0.19623	1.21	<0.094
Among three altitude groups	2	58.114	29.06	-0.04179	0.26	< 0.535
Among populations	5	141.008	28.2016	1.10478	6.79	<0.000
Within population	88	1316.714	14.963	14.962	92.00	<0.000

separated on 5% denaturing polyacrylamide sequencing gels (19:1 acrylamide: bis-acrylamide, 7 M urea) in 1X TBE buffer (1 M Tris, 1 M boric acid, 20 mM EDTA). The gels were pre-run for 40 min before 3  $\mu$ l of the mix was loaded and run at 85 W for 3 h. Bands were visualised in sodium carbonate developing solution for 5 min and silver-stained gels were rinsed in ultra pure water and dried at room temperature in fumehood.

#### Data analysis

AFLP bands between 100 and 550 bp were scored twice manually and independently from the glass plates and counter checked. Polymorphic AFLP fragments were scored as 1 and 0 for presence and absence of bands, respectively, and data were entered into a binary matrix for each marker as discrete variables. The binary matrices were analysed by the software Arlequin 2.0 (Schneider et al., 2000) for calculating average difference between all the genotypes in the population (Tajima, 1993) and average gene diversity over loci (Nei, 1987). For each primer pair, the numbers of polymorphic and monomorphic bands were determined; however, bands that were monomorphic across all the 96 individuals were excluded from analyses. A matrix of corrected average pairwise differences between all pairs of populations generated by Arlequin was used for a constructing dendrograms through cluster analysis using the unweighted pair group method based on arithmetic averages (UPGMA) in the programme NTSYS-pc version 2.1 (Rohlf, 2000). Simple Matching and Jaccard's coefficients of similarity (Sneath and Sokal, 1973) were calculated for all pairwise comparisons among populations and the coefficients presented similar results (only used the analyses based on Jaccard's coefficients were used). Unscrambler 9.5 from CAMO in Oslo was used to investigate further relationships among the populations

using principal component analysis (PCA) to show clusters of relatedness among *U. kirkiana* populations.

## RESULTS

### Level of polymorphism

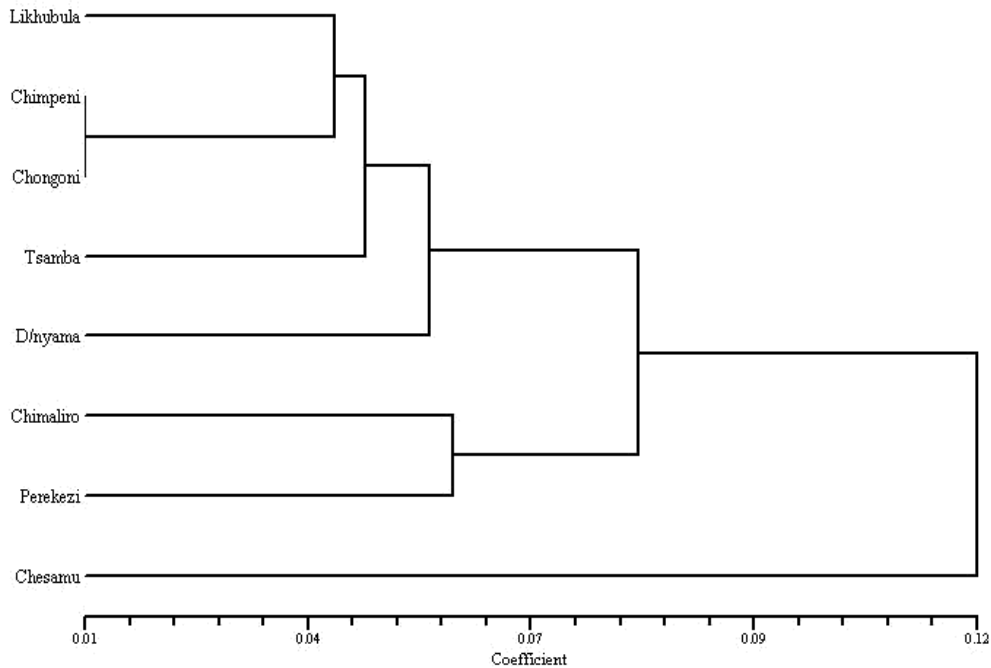
The six primer combinations generated a total of 612 AFLP bands out of which 110 were polymorphic representing 18% polymorphism. Total number of polymorphic loci scored per primer pair ranged from 10 (E-AAG/M-CTA) to 32 (E41+A/MCTA) with an average of 18 loci per primer combination (Table 2) and these numbers suggest a low degree of variability.

### Genetic diversity and differentiation

Analysis of molecular variance (AMOVA) (Excoffier et al., 1992) revealed low but significant genetic variation among populations of *U. kirkiana* but there were no significant genetic variations among *U. kirkiana* in the three regions and between the altitudes (low, medium and high) (Table 3). The mean Nei's (1973) genetic diversity for all the 110 loci in the 8 populations was  $0.27 \pm 0.141$  and the mean Nei's genetic diversity for populations ranged from  $H=0.223$  to  $H=0.322$ . Tsamba population had the lowest average gene diversity

**Table 4.** Genetic diversity indices in 8 Malawian *Uapaca kirkiana* populations. The indices calculated are frequency of polymorphic loci, mean number of pairwise differences between individuals and average gene diversity over loci.

Population	Frequency of polymorphic loci	Average difference	Average gene diversity (H)
Likhubula	0.82	34.74	0.322
Chimpeni	0.76	29.02	0.279
Tsamba	0.61	23.89	0.223
Chongoni	0.76	28.41	0.268
Dzalanyama	0.69	22.82	0.240
Chimaliro	0.78	29.35	0.282
Perekezi	0.76	27.23	0.279
Chesamu	0.78	27.23	0.269
Average	0.75 ± 0.08	27.84 ± 12.01	0.270 ± 0.14



**Figure 2.** Dendrogram of 8 populations using 110 polymorphic AFLPs based on UPGMA of corrected average pairwise differences.

whereas Likhubula had the highest average gene diversity (Table 4).

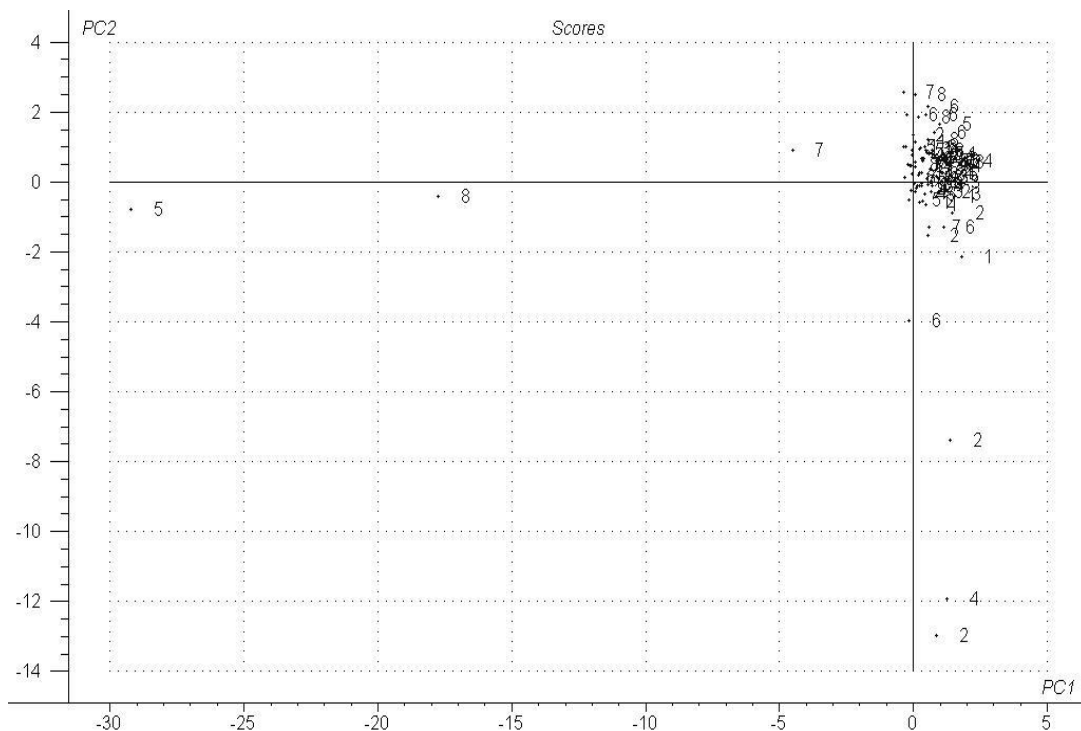
### Cluster analyses

The dendrogram generated from AFLP data using UPGMA is shown in Figure 2. There is no clear structuring of the variability relative to region, however three clusters in UPGMA can be defined by cutting the dendrogram at a genetic similarity value of 0.078 and the populations separate according to geographic origin with first cluster composed of 3 populations from southern Malawi (Likhubula, Chimpeni and Tsamba) and two from

Central Malawi populations (Chongoni and Dzalanyama). The second cluster is composed of Chimaliro from the Central region which clustered with Northern region population of Perekezi leaving Chesamu from Northern region as a third separate cluster.

### Principal component analysis

Results of principal component analysis (Figure 3) show that only 36% of the total variation can be explained by the three PCA based on first-second and third eigenvectors which account for 22, 8 and 6%, respectively. The eight populations fall in one cluster although some individuals from populations 2, 4, 5 and 8



**Figure 3.** Principal component analyses of populations of *Uapaca kirkiana* using 110 AFLP polymorphic markers.

were separated from the cluster. Excluding the six individuals, the analysis did not show separation of the eight populations into different clusters (data not shown).

## DISCUSSION

The data exhibit very high genetic diversity within population compared to among population of *U. kirkiana*; this is in agreement with its life history traits and geographical distribution. Very little is known about the mating system of *U. kirkiana*, however, work by Ngulube et al. (1998) has shown that the species is highly out crossing. High within population variability is expected from such a plant with wide and continuous distribution (Hamrick and Godt, 1989). The highest levels of genetic diversity ( $H=0.322$ ) was found within the broader distribution of Likhubula population in Southern Malawi with the restricted population of Tsamba in the same region showing lowest level of diversity. Distribution range and population size have been identified as major correlates of within population genetic variation in tropical tree species with restricted populations showing significantly less variation than those with a broader distribution (Loveless, 1992). In general, populations collected from protected forest reserves had higher genetic diversity than populations collected from customary and private forest suggesting negative impact

of human activities on genetic diversity. This is consistent with field observations that the numbers of *U. kirkiana* trees and saplings in forest reserves were higher than on customary and private forests (Mwase et al., in preparation).

The results based on AMOVA show strong intrapopulation variation with small differentiation among the eight populations. The results agree with the general observation that woody, perennial, outbreeding species maintain most of their variation within populations (Hamrick et al., 1992). The lack of differentiation among regions and different altitude is indicative of high levels of gene flow and this is typical of woody outcrossing species. The low, medium and high altitude areas are randomly distributed in Malawi and they are often adjacent to each other in which case gene flow would be high. The negative variance among regional samples implies that alleles are more related than within populations. The lack of differentiation is also indicative of lack of sufficient time for significant differentiation along geographic lines. *U. kirkiana* is a pioneer plant species growing in open canopies in miombo woodland and low population differentiation can also be attributed to its longevity traits which are not very typical of early successional species. According to Wright's (1978) guidelines for interpretation of  $F_{ST}$  values, the range 0.0-0.005 indicates little, 0.05-0.15 moderate, 0.15-0.25 large and >0.25 very large population differentiation. The value

of  $F_{ST}$  0.079 suggests moderate differentiation and is consistent with the life history of *U. kirkiana*. Tropical trees tend to possess the most genetic diversity within populations (Hamrick and Loveless, 1989), and species reproducing both sexually and asexually show less differentiation than species reproducing only sexually (Hamrick and Godt, 1989). Although *U. kirkiana* reproduces both sexually and vegetatively by sprouting of prostrate stems; this can be viewed as a strategy for reproductive success which is common in woody plants exposed to environmental conditions. The results agree with Agufa (2000) who reported high genetic diversity within populations of 91% for *U. kirkiana* samples collected from Zambia, Malawi, Zimbabwe and Tanzania with very low diversity among countries. The values are similar to those obtained in other studies of outcrossing tree species; for example, *Swietenia macrophylla* (Gillies et al., 1999) and *Populus tremuloides* (Yeh et al., 1995) where 87.9 and 97.4% of their genetic variation resided within populations, respectively. Lack of genetic variation among the three geographical regions suggests considerable gene flow due to exchange of plant material taking place over longer distances of more than 200 km. Geographical distances between populations vary from 120 to 800 km. Natural seed dispersal by rodents and birds are thought to take place for very longer distances of up to 50 km. Taking into account the biological features of the plants, high effective gene flow would be expected. Human transportation of fresh fruits across large distances between forests and towns might have improved genetic exchange through seed dispersal within species natural distribution. Farmers selling fruits by road side markets and at markets in the towns may be responsible for long distance seed dispersal. The extensive gene flow could to a certain extent counteract effects of genetic drift in populations in the different geographical regions and act as a homogenization factor between nearby populations.

The low percentage (36%) of the variation explained by the first three components in the PCA could be attributed to high extent of genetic variability between individuals of each population than the low variation among the populations. The PCA support the AMOVA analysis. The PCA results analysis shows that the 8 populations did not segregate into distinguishable clusters; this is consistent with AMOVA results which indicated that genetic diversity was higher between individuals within a population than between populations. Even though the genetic differentiation among populations was significant, it accounted for only 6.8% of the variation. However the PCA showed two individuals from populations 2 (Chimpeni) and 4 (Chongoni) clustering together and this agrees with results of the UPGMA but the clustering by PCA did not agree in its entirety with UGMA. This could be attributed to the sensitivity of the PCA since this analysis is expected to be more informative about differentiation among major groups while the cluster

analysis of UPGMA provides resolution among closely related populations. Populations tended to cluster together with other populations from the same region. However, Chimaliro from central region clustered with northern population of Perekezi indicating relatively high geographic proximity of the two populations. The two populations, even though belong to different regions, are close to each other and this facilitates gene exchange. Chesamu population is located on a physically isolated lakeshore area and the isolation may have influenced its genetic diversity through genetic drift.

Populations from communal forests of Tsamba had the lowest genetic diversity probably due to a founder effect since this is a population of recent origin. This result agrees with the prediction that marginal populations should be less variable than populations in the primary range (Stewart and Nilsen, 1995). Low genetic diversity in this low altitude region could also be attributed to extensive exploitation of the trees species by local communities. Changes in the landscape caused by deforestation and habitat destruction alter population density, abundance, diversity and abundance of pollinator communities (Lowe et al., 2005) thereby impinging on the mating system. The results signify the importance of focusing on populations which show more variability for conservation purposes. The present data pattern suggests that Likhubula population should be considered more distinct for the purposes of seed collection and management. However, for the purpose of conservation of genetic resources seed collection ought to be done across different populations to ensure a more representative sample of genetic variation. The country has shown low levels of genetic variation among the populations but very high within population. The results have some implications for conservation of *U. kirkiana* in Malawi. When selecting trees for clonal propagation, there is need to maximise selection from individual populations further suggesting that germplasm evaluation and conservation of *U. kirkiana* should focus on extensive sampling of more individuals especially from populations that are genetically diverse.

## CONCLUSION

The analysis of AFLP exhibited very low genetic variation among *U. kirkiana* populations but very high variation between individuals in a population characteristic of an outbreeding species. These results are of great relevance for domestication and conservation of *U. kirkiana*. Priority on conservation and collection of germplasm should be given to populations with higher genetic diversity and it would be cost effective to sample a small number of populations in a region represented by a large number of individuals from a population. The genetic structure of the eight populations appears within the same range with higher genetic diversity values for population from forest

reserves than populations from customary land. Overall, the study has demonstrated usefulness of AFLP markers in detecting the genetic diversity over a large number of randomly sampled loci and it has proved its usefulness in discriminating *U. kirkiana* populations and individuals within a population. Finally, our results indicate that a more intense monitoring of genetic diversity using AFLP should be conducted for *U. kirkiana* provenances in East and Southern African countries to identify candidate populations for conservation among the countries.

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## REFERENCES

- Agufa CAC (2002). Genetic variation in *Sclerocarya birrea* and *Uapaca kirkiana* -indigenous fruit trees of the miombo woodlands. MSc thesis Jomo Kenyatta University of Agriculture and Technology, Nairobi Kenya.
- Excoffier L, Smouse PE, Quattro JM (1992). Analysis of molecular variance inferred from metric distances among DNA haplotypes: application to human mitochondrial DNA restriction data. *Genetics* 131: 179-191.
- Gillies ACM, Navarro C, Lowe AJ, Newton AC, Hernandez M, Wilson J, Cornelious JP (1999). Genetic diversity in Mesoamerican populations of mahogany (*Swietenia macrophylla*) assessed using RAPD. *Heredity* 83: 722-732.
- Hamrick JL, Godt MJW (1989). Allozyme diversity in plant species. In: *Genetics and Conservation of Rare plants*, eds Falk DD, Holsinger, KE. Oxford University Press, Oxford, pp. 75-86.
- Hamrick JL, Godt MJ, Sherman-Broyles SL (1992). Factors influencing levels of genetic diversity in woody plant species. *New Forest* 6: 95-124.
- Hamrick JL, Loveless MD (1989). The genetic structure of tropical tree populations: associations with reproductive biology. In: Bock JH, Linhart YB (eds). *The evolutionary ecology of plants*. Boulder, CO, Westview Press. pp. 129-146.
- Hamrick JL, Godt MJW, Murawski DA, Loveless MD (1991). Correlations between species trait and allozyme diversity: Implications for conservation biology. In: *Genetics and Conservation of Rare plants* (eds) Falk DD, Holsinger KE) pp 75-86. Oxford University Press, Oxford.
- Heckenberger M, vander Voort J, Melchinger AE, Peleman J, Bohn M (2003). Variation in DNA fingerprints among accessions with maize inbred lines and implications for identification of essentially derived varieties II. Genetic and technical sources of variation in AFLP data and comparison to SSR data. *Mol. Breed.* 12: 97-106.
- Karachi M, Ruffo CK, Lema NM, Minae S (1991). Use of multipurpose trees in western Tanzania: results of an ethnobotanical survey of multipurpose trees in the Tabora Region. *AFRENA Report No. 40*. Int. Centre for Res. in Agroforestry, Nairobi, Kenya.
- Kwesiga F, Chisumpa SM (1992). Multipurpose trees for the Eastern Province of Zambia: an ethnobotanical survey of their use in the farming systems. *AFRENA Report No. 49*. Int. Centre for Res. in Agroforestry, Nairobi, Kenya.
- Loveless MD (1992). Isozyme variation in tropical trees: patterns of genetic organization. *New Forests* 6: 67-94.
- Maghembe JA, Seyani JH (1992). Multipurpose trees used by small-scale farmers in Malawi: results of an ethnobotanical survey. *AFRENA Report No. 42*. Int. Centre for Res. in Agroforestry, Nairobi, Kenya.
- Maghembe JA, Kwesiga F, Ngulube M, Prins H, Malaya F (1994). Domestication potential of indigenous fruit trees of the Miombo Woodlands of Southern Africa. In: Leakey, R.R.B and Newton, A.C. (eds.) *Tropical trees: Potential for Domestication and the Rebuilding of Forest Resources*, pp. 220-229. HMSO, London, UK.
- Marghali S, Panaud O, Lamy F, Ghariani S, Sarr A, Marrakchi M, Trifi-Farah N (2005). Exploration of intra- and inter-population genetic diversity in *Hedysarum coronarium*, L. by AFLP markers. *Genetic Resources and Crop Evolution* 8: 277-284.
- Mwamba CK (1995). Variations in fruits of *Uapaca kirkiana* and effects of in situ silvicultural treatments on fruit parameters: In *Improvement of indigenous fruit trees of the miombo woodlands of southern Africa*, pp. 50-57. Int. Centre for Res. in Agroforestry, Nairobi, Kenya. Muluvi GM, Sprent JI, Soranzo N, Provan J, Odee D, Folkard G, McNiccol JW, Powell W (1999). Amplified Fragment Length Polymorphism (AFLP) analysis of genetic variation in *Moringa oleifera* Lam. *Molecular Ecol.* 8: 463-470.
- Nei M (1987). *Molecular Evolutionary Genetics*. Columbia University Press, New, NY, USA
- Ngulube MR, Hall JB, Maghembe JA (1995). Ecology of a miombo fruit tree: *Uapaca kirkiana* (Euphorbiaceae). *Forest Ecol. Management* 77: 107-111.
- Ngulube MR, Kanaji B. (1989). Seed problems of edible indigenous fruits in Malawi. In: Kwapata, M.B, Kanyama Phiri, G.Y and Kasomekera, Z, (eds). *Proceedings of the iiiith Annual Conference on Science and technology for development, AASToM held in Blantyre*, 104-109.
- Ngulube MR, Hall JB, Maghembe JA (1997). Fruit, seed and seedling variation in *Uapaca kirkiana* from natural populations in Malawi. *For. Ecol. Management* 98: 209-219.
- Ngulube MR, Hall JB, Maghembe JA (1998). Reproductive ecology of *Uapaca kirkiana* (Euphorbiaceae) in Malawi, southern Africa. *J. Tropical Ecol.* 14: 743-760.
- Paterson AH, Brubaker CL, Wendel JF (1993). A Rapid method for extraction of cotton (*Gossypium* species) genomic DNA suitable for RFLP or PCR analysis. *Plant Mol. Biol Rep* 11: 122-127.
- Radcliffe-Smith A (1988). Euphorbiaceae (part 2): *Uapaca*. In: Polhill, R.M. (ed.) *Flora of Tropical East Africa*. Balkema, Rotterdam/Brookfield.
- Rohlf FJ (2000). *NTSYS-pc. Numerical Taxonomy and Multivariate analysis system*, version 2.1. Exeter Software, New York, USA.
- Sambrook J, Fritsch EF, Maniatis T (1989). *Molecular cloning: a laboratory manual*. 3<sup>rd</sup> edition. Cold Spring Harbor Laboratory Press, New York.
- Schmidt K, Jenssen K (2000). Genetic structure and AFLP variation of remnant populations in the rare plant *Pedicularis palustris* (Scrophulariaceae) and its relation to population size and reproduction components. *American Journal of Botany* 87: 678-689.
- Schneider S, Roessli D, Excoffier L, (2000). Arlequin version 2.0: A software for population genetics data analysis. *Genetics and Biometry Laboratory*, University of Geneva, Switzerland.
- Sneath PHA, Sokal RR (1973). *Numerical Taxonomy*. Freeman, San Francisco, USA.
- Stewart CN, Nilsen ET (1995). Phenotypic plasticity and genetic variation in *Vaccinium macrocarpon*, the America cranberry. I. Reaction norms of clones from central and marginal populations in a common garden. *Int. J. Plant Sci.* 156:687-697.
- Tajima F (1993). Measurement of DNA polymorphism. In: Takahata N, Clark AG. (eds). *Mechanisms of Molecular Evolution. Introduction to Molecular Paleopopulation Biology*. Sinauer associates, Sunderland, MA, pp 37-59.
- Vos P, Hogers R, Bleeker M, Reijmans M, van der Lee T, Hornes M, Frijters A, Pot J, Peleman J, Kuiper M, Zabeau M (1995). AFLP: A new



technique for DNA fingerprinting. *Nucleic Acids Res.* 23: 4407-4414.  
Wright S (1978). *Evolution and genetics of populations: Variability within and among natural populations*. Vol. 4, University of Chicago Press, Chicago, USA.

Yeh FC, Chong DKK, Yang RC (1995). RAPD variation within and among natural populations of trembling aspen (*Populus tremuloides*) from Alberta. *J. Heredity* 86: 454-460.