

Full Length Research Paper

# Phytochemical constituents and antioxidant properties of extracts from the leaves of *Chromolaena odorata*

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Aqueous and methanolic extracts of *Chromolaena odorata* were screened for phytochemical constituents. The evaluation of the antioxidant potential of the methanolic extract was also carried out. Tests for tannins, steroids, terpenoids, flavonoids and cardiac glycosides were positive in both methanolic and aqueous extracts. Alkaloids were detected only in the methanolic extract. The total phenolic content, reducing power and percent DPPH scavenging effect were  $0.01 \pm 0.00$  mg/g GAE,  $0.22 \pm 0.01$  and  $28.85 \pm 0.99\%$ , respectively. Against the backdrop of many known medicinal properties of this plant, results from the present work suggest that relatively low values of antioxidant indices may not imply a low medicinal value.

Key words: Chromolaena odorata, phytochemicals, antioxidant activity, reductive potential, medicinal plants

# INTRODUCTION

Chromolaena odorata (L) King and Robinson Asteraceae commonly known as Siam weed, is a fast-growing pere-nnial and invasive weed native to South and Central America. It has been introduced into the tropical regions of Asia, Africa and other parts of the world. It is an aggre-ssive competitor that occupies different types of lands where it forms dense strands that prevents the establish-ment of other flora. It is a menace in plantations and other ecosystems. It suppresses young plantations, agri-cultural crops and smothers vegetation as it possesses allelopathic potentialities and growth inhibitors (Ambika and Jayachandra, 1980; Ambika and Jayachandra, 1982; Muniappan and Marutani, 1988). The economic value of C. odorata is low. Consequently, there is a relative pau-city of research works on it. It is a perennial shrub native of South and Central America. In recent decades, it has become a serious pest in the humid tropics of South East Asia, Africa and Pacific Islands. It spreads rapidly in lands used for forestry, pasture and plantation crops such as rubber, coffee, coconut, cocoa and cashew. The plant can be poisonous to livestock as it has exceptionally high level of nitrate (5 to 6 times above the toxic level) in the

leaves and young shoots; the cattle feeding on these die of tissue anoxia (Sajise et al., 1974).

Despite the negative sides to the plant, it still has patronage from practitioners of traditional medicine. It has been reported to have antispasmodic, aniprotozoal, antitrypanosomal, antibacterial and antihypertensive activeties. It has also been reported to possess anti-inflammatory, astringent, diuretic and hepatotropic activities (Watt and Brandwijk, 1962; Feng et al, 1964; Weniger and Robinean, 1988; Iwu, 1993). In the southern part of Nigeria, the leaves are used for wound dressing, skin infection and to stop bleeding. Some specific phenolic compounds have been isolated from the plant (Metwally and Ekejuba, 1981).

The medicinal values of plants lie in their component phytochemicals such as alkaloids, tannins, flavonoids and other phenolic compounds, which produce a definite physiological action on the human body (Hill, 1952). A systematic search for useful bioactivities from medicinal plants is now considered to be a rational approach in nutraceutical and drug research. Therefore, the present work has been designed to evaluate the antioxidant potential of *C. odorata* with a view to contributing to the search for beneficial uses of this invasive plant which is a menace to farmers.

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Phytochemicals	Methanolic extracts	Aqueous extracts
Alkaloids	+	-
Saponins	-	+
Tannins	+	+
Phlobatannins	+	+
Anthraquinones	-	+
Steroids	+	+
Terpenoids	+	+
Flavonoids	+	+
Cardiac glycosides		
With steroidal ring	+	+
With deoxy – sugar	+	+

Table 1. Phytochemicals detected in extracts of Chromolaena odorata.

+ = Present

- = Absent

#### MATERIALS AND METHODS

#### Chemicals

DPPH (2, 2-diphenyl-1-picrylhydrazyl) radical, gallic acid, ascorbic acid and Folin-Ciocalteau reagent were obtained from Sigma-Aldrich, USA. All other chemicals and reagents used were of analytical grade.

#### **Plant materials**

Leaves of *C. odoarata* were collected from a farmland in Akure, South-Western Nigeria and identified at the Department of Crop, Soil and Pest Management, Federal University of Technology, Akure. They were air dried, packed in paper bags and stored. The dried leaves were pulverized and 196 g of the pulverized sample was extracted with 500 ml of 80% methanol by maceration for 72 h. The methanolic extract was concentrated in a rotary evaporator, lyophilized and thereafter preserved for further use. An aqueous extract was also prepared from the pulverized sample for the purpose of comparison of the phytochemical constituents with that of the methanolic extract.

#### Phytochemical screening

Chemical tests were carried out on the aqueous and methanolic extracts for the qualitative determination of phytochemical constituents as described by Harborne (1973), Trease and Evans (1989) and Sofowora (1993). Total phenolic content was determined using Folin-Ciocalteau reagent as previously described (McDonald et al., 2001). Total phenol value was expressed as mg/g gallic acid equivalent.

#### **DPPH radical scavenging activity**

The ability of the extract to scavenge DPPH radical was determined according to the method described by Mensor et al. (2001). One ml of a 0.3 mM DPPH methanol solution was added to a solution of the extract or standard (250  $\mu$ g/ml, 2.5 ml) and allowed to react at room temperature for 30 min. The absorbance of the resulting mixture was measured at 518 nm and converted to percentage antioxidant activity (AA %). Methanol (1.0 ml) plus extract solution (2.5 ml) was

used as a blank. 1 ml of 0.3 mM DPPH plus methanol (2.5 ml) was used as a negative control. Solution of gallic acid served as positive control.

#### **Reductive potential**

This was determined according to the method of Oyaizu (1986). The extract or standard (100  $\mu$ g/ml) was mixed with phosphate buffer and potassium ferricyanide. The mixture was incubated at 50°C for 20 min. Tricloroacetic acid (10%, 2.5 ml) was added to the mixture. A portion of the resulting mixture was mixed with FeCl<sub>3</sub> (0.1%, 0.5 ml) and the absorbance was measured at 700 nm in a spectrophotometer. Higher absorbance of the reaction mixture indicates higher reductive potential.

#### Statistical analysis

Data were expressed as mean  $\pm$  SEM. A one-way analysis of variance was used to analyze data. P<0.5 represented significant difference between means (Duncan's multiple range test).

## **RESULTS AND DISCUSSION**

Table 1 shows the phytochemicals detected in *C. odorata* leaf extract. Tests for tannins, steroids, terpenoids, flavornoids and cardiac glycosides were positive in both methanolic and aqueous extracts. Alkaloids were detected only in the methanolic extract. Phenolics, alkaloids, terpenoids and cardiac glycosides detected in the extracts are compounds that have been documented to possess medicinal properties and health-promoting effects (Salah et al., 1995; Del-Rio et al., 1997; Okwu, 2004; Liu, 2004). The total phenolic content in the methanolic extract was  $0.01 \pm 0.00 \text{ mg/gGAE}$ ; a rather low value. Phenolics are the largest group of phytochemicals and have been said to account for most of the antioxidant activity of plant extracts (Thabrew et al., 1998).

The result of the DPPH scavenging assay is shown in Figure 1. The percentage antioxidant activity of *C. odora*-



**Figure 1.** Antioxidant activity of CO compared with some standards. CO, *Chromolaena odorata*; GA, gallic acid; and AA, ascorbic acid. <sup>#</sup>Not significantly different, P>0.05 and \*Significantly different, P<0.001.



**Figure 2.** Reductive potential of *Chromolaena odorata*. AA, Ascorbic acid; and CO, *Chromolaena odorata*. \*<sup>#</sup> Significantly different (P<0.001).

*ta* is about a third of the value for gallic acid and ascorbic acid while the reductive potential is about a fifth of the value for ascorbic acid. These values are appreciable enough for a plant that has been tagged as an obnoxious weed.

The relatively low values for the percentage antioxidant activity (Figure 1) and the reductive potential (Figure 2) are in harmony with the finding that a strong positive association exists between total phenolic content and DPPH scavenging effect and also between total phenolic content and reductive potential (Miliauskas et al., 2004) ). Low values in some antioxidant assays do not imply low value in all other assays since many antioxidant assays show no correlation (Schaich, 2006). Result of a particular antioxidant assay depends on the chemistry of the assay and the nature and combination of bioactive princi-

ples in the material under investigation.

Emerging trends in antioxidant research point to the fact that low levels of phenolics (and other phytochemicals) and low values of antioxidant indices in plants do not translate to poor medicinal properties. Mineral elements, other secondary plant metabolites not detected or evaluated and vitamins contribute to the synergy of phytochemicals that confer medicinal properties on plants. The present investigation indicates that though C. odorata has been described as a plant of low economic value, it is not worthless. Its use in traditional medicine attests to this. There are prospects for its commercial utilization especially in view of its abundant and widespread nature. The toxic compounds in the plant could be removed through appropriate extraction and processing methods making extracts and products from the plant safe for the utilization of animal and man. Control of this invasive and notorious weed has been unsuccessful. Finding ways of profitably utilizing it may be the best option left. Further work is in progress in our laboratory along these lines.

#### REFERENCES

- Ambica SR Jayachandra (1980). Suppression of plantations crops by Eupatorium weed. Curr. Sci. 49: 874-875.
- Ambica SR Jayachandra (1982). Eupatorium odoratum L. in plantations – An allelopath or a growth promoter? "In proceedings of the fifth annual symposium on plantation crops, held at CPCRI, Kasaragog, Dec 15-18.
- Del-Rio A Obdulio BG, Castillo J, Marin RR, Ortuno A (1977). Uses and properties of citrus flavonoids. J. Agric. Food Chem. 45: 4505-4515.
- Feng PC, Haynes LJ, Magnues KE, Plimmer JR (1964). Further pharmacological screening of some West Indian medicinal plants. J. Pharm. Pharmacol. 16: 115-117.
- Harborne JB (1973). Phytochemical methods. Chapman and Hall Ltd., London. pp. 49-188.
- Hill AF (1952). Economic Botany. A textbook of useful plants and plant products. 2<sup>nd</sup> edn. McGraw-Hill Book Company Inc, New York.
- Iwu MM (1993). Handbook of African Medicinal Plants, CRC Press Inc., Beca Raton. pp. 181-182.

- Liu RH (2004). Potential synergy of phytochemicals in cancer prevention: mechanism of action. J. Nutr., 134: 3479S-3485S.
- McDonald S, Prenzler PD, Autolovich M, Robards K (2001). Phenolic content and antioxidant activity of olive extracts. Food Chem. 73: 73-84.
- Mensor LI, Menezes FS, Leitao GG, Reis AS, dos Santos T, Coube CS Leitao, SG (2001). Screening of Brazillian plant extracts for antioxidant activity by the use of DPPH free radical method. Phytother. Res. 15: 127-130.
- Metwally AM Ekejuba EC (1981). Methoxylated flavonols and flavones from *Eupatorium odoratuml*. Planta Medica 42: 403.
- Miliauskas G, Venskutonis PR, vanBeek TA (2004). Screening of radical scavenging activity of some medicinal and aromatic plant extracts. Food Chem. 85: 231-237
- .Muniappan R, Marutani M (1998). Ecology and distribution of C. odorata in Asia and Pacific. In the Proceedings of the First International Workshop on Biological Control of C. odorata held from Feb 29-Mar 4, Bangkok, Thailand.
- Okwu DE (2004). Phytochemical and vitamin content of indigenous spices of South Eastern Nigeria. J. Sustain. Agric. Environ., 6: 30-34.
- Oyaizu M (1986). Studies on product of browning reaction prepared from glucose amine. Jpn. J. Nutr. 44: 307-315.
- Salah W, Miller NJ, Pagauga G, Tijburg, Bolwell GP, Rice E, Evans C (1995). Polyphenolic flavonols as scavenger of aqueous phase radicals and chainbreaking antioxidants. Arch. Biochem. Biol. 2: 339-346.
- Sajise PE, Palis RK, Norcio NV, Lales JS (1974). The biology of *C. odorata* L. King and Robinson. 1. Flowering behaviour, pattern of growth and nitrate metabolism. Phil. Weed Sci. Bull. 1:17-24.
- Schaich KM (2006). Developing a rational basis for the selection of antioxidant screening and testing methods. ISHS Acta Horticulturae p. 709.
- Sofowora LA (1993). Medicinal plants and traditional medicine in Africa. Spectrum Books Ltd, Ibaban. Harborne. pp. 55-71.
- Taiwo OB, Olajide OA, Soyannwo OO, Makinde JM (2000). Antiinflammatory, antipyretic and antispasmodic properties of *Chrom*olaena odorata. Pharm. Biol. 38: 367-370.
- Thabrew MI, Hughes RD, McFarlane IG (1998). Antioxidant activity of *Osbeckia aspera*. Phytother. Res. 12: 288-290.
- Trease GE, Evans WC (1989). Trease and Evans' Pharmacognosy: A Physician's Guide to Herbal Medicine. 13th Edition, Bailliere Tindall London.
- Watt JM, Breyer-brandwijik MG (1962). Medicinal and Poisonous Plants of Southern and Eastern Africa. E and S Livingstone, Edinburgh.
- Wniger B Robinean L (1988). Elements for Carribean Pharmacopoiea. Proceedings of TRAMIL workshop, Cuba.